

Increased phosphorus availability mitigates the inhibition of nitrogen deposition on CH₄ uptake in an old-growth tropical forest, southern China

T. Zhang^{1,3}, W. Zhu², J. Mo¹, L. Liu^{1,3}, and S. Dong¹

¹Key Laboratory of Vegetation Restoration and Management of Degraded Ecosystems, South China Botanical Garden, Chinese Academy of Sciences, Guangzhou 510650, China

²Department of Biological Science, State University of New York – Binghamton, Binghamton NY, 13902, USA

³Graduate University of Chinese Academy of Sciences, Beijing 100039, China

Received: 21 March 2011 – Published in Biogeosciences Discuss.: 23 May 2011

Revised: 5 September 2011 – Accepted: 13 September 2011 – Published: 29 September 2011

Abstract. It is well established that tropical forest ecosystems are often limited by phosphorus (P) availability, and elevated atmospheric nitrogen (N) deposition may further enhance such P limitation. However, it is uncertain whether P availability would affect soil fluxes of greenhouse gases, such as methane (CH₄) uptake, and how P interacts with N deposition. We examine the effects of N and P additions on soil CH₄ uptake in an N saturated old-growth tropical forest in southern China to test the following hypotheses: (1) P addition would increase CH₄ uptake; (2) N addition would decrease CH₄ uptake; and (3) P addition would mitigate the inhibitive effect of N addition on soil CH₄ uptake. Four treatments were conducted at the following levels from February 2007 to October 2009: control, N-addition (150 kg N ha⁻¹ yr⁻¹), P-addition (150 kg P ha⁻¹ yr⁻¹), and NP-addition (150 kg N ha⁻¹ yr⁻¹ plus 150 kg P ha⁻¹ yr⁻¹). Static chamber and gas chromatography techniques were used to quantify soil CH₄ uptake every month throughout the study period. Average CH₄ uptake rate was 31.2 ± 1.1 μg CH₄-C m⁻² h⁻¹ in the control plots. The mean CH₄ uptake rate in the N-addition plots was 23.6 ± 0.9 μg CH₄-C m⁻² h⁻¹, significantly lower than that in the controls. P-addition however, significantly increased CH₄ uptake by 24 % (38.8 ± 1.3 μg CH₄-C m⁻² h⁻¹), whereas NP-addition (33.6 ± 1.0 μg CH₄-C m⁻² h⁻¹) was not statistically different from the control. Our results suggest that increased P availability may enhance soil methanotrophic activity and root growth, resulting in potentially mitigating the inhibitive effect of N deposition on CH₄ uptake in tropical forests.

1 Introduction

Methane (CH₄) is considered the second most important greenhouse gas after carbon dioxide and with a global warming potential 25 times compare to carbon dioxide (CO₂) over a 100 year horizon (IPCC, 2007). The global atmospheric concentration of CH₄ has increased from a pre-industrial value of about 0.715 ppm to 1.732 ppm in the early 1990s, and 1.803 ppm in 2009 (IPCC, 2007; WMO, 2010), due primarily to the anthropogenic emissions from energy production, rice cultivation, ruminant animals, biomass burning and landfills (IPCC, 2007).

Major CH₄ sinks include tropospheric oxidation (~500 Tg yr⁻¹) and stratospheric loss (~40 Tg yr⁻¹) (Hein et al., 1997; Lelieveld et al., 1998). Upland oxic soil that are continuously emerged and exposed to atmospheric concentrations of CH₄ is another major sink of atmospheric CH₄, which consumes CH₄ through the activity of methanotrophs under aerobic conditions (~30 Tg CH₄ yr⁻¹) (Lelieveld et al., 1998). Among upland soils, forest soils are probably the most efficient CH₄ sink (Le Mer and Roger, 2001).

Atmospheric methane originates mainly from a biological process. In forest soils, it is produced in anoxic layer by methanogenic bacteria during the anaerobic digesting of organic matter. Methane is also eliminated into forest soils by microbial oxidation (methanotrophy) in the aerobic zone and which oxidises atmospheric methane. Methanotrophs in forest soil use CH₄ as only a carbon (C) and energy source and oxygen availability is the main factor limiting their activity (Le Mer and Roger, 2001). Therefore, methanotrophy in forest soils is primarily dependent on physical factors controlling soil diffusion, e.g., water content and soil texture (Butterbach-Bahl and Pappen, 2002; Templeton et al.,



Correspondence to: J. Mo
(mojm@scib.ac.cn)

2006). Teh et al. (2005) indicate that the effective diffusion at soil surface controls the overall rate of methane consumption. However, forest soils as biological sinks of CH₄ are also subjected to other abiotic and biotic controls. It is well known that methanotrophs have different temperature and pH optima. Effect soil temperature on methanotrophic community has been well studied (Mohanty et al., 2007). Soil pH seems to be a less important control for atmospheric methane oxidation (Kolb, 2009). Tree species would affect atmospheric CH₄ oxidation without altering community composition of soil methanotrophs (Menyailo et al., 2010), Jamali et al. (2011) found that termites offset 21 % of CH₄ consumed by soil in a tropical savanna woodland. Furthermore, it is believed that nitrogen (N) availability controls the activity and affects the community structure of methanotrophs in soil (Bodelier and Laanbroek, 2004).

Among the above factors, the effects of nitrogen deposition on CH₄ oxidation have received increasing attention (Stuedler et al., 1989; Bodelier and Laanbroek, 2004; Tate et al., 2007; Zhang et al., 2008). Nitrogen addition alters the fluxes of greenhouse gases (GHGs, including CH₄) through regulating plant and microbial activities that are directly associated with GHGs production and consumption (Liu and Greaver, 2009). Stuedler et al. (1989) first reported in a temperate forest that N fertilization reduced soil CH₄ uptake by 33 %. Extensive research has been conducted to investigate the relationship between CH₄ consumption and N input and it has been generally accepted that CH₄ uptake is inhibited by nitrogenous fertilization (Bodelier and Laanbroek, 2004; Chan et al., 2005). Laboratory studies indicate that oxidation of CH₄ by a variety of methanotrophs is competitively inhibited by N (Ferenci et al., 1975). In soils, the inhibition of CH₄ oxidation by ammonium is attributed to a competition at the level of the methane mono-oxygenase, a transfer of the methanotrophy activity towards nitrification (Castro et al., 1994) and the toxicity of NO₂ produced. Aluminium toxicity after extensive N input may also inhibit soil CH₄ uptake (Bradford et al., 2001a; Zhang et al., 2008). On the other hand, positive effects of increased N availability on CH₄ uptake rates were found in severely N-limited forests (Börjesson and Nohrstedt, 2000; Steinkamp et al., 2001). In addition, soil available phosphorus would also affect CH₄ flux by regulating changes in soil physico-chemical properties, plant root activities and soil microbial activities that are involved in CH₄ consumption. Increased P availability in forest soil would stimulate plant root growth and lead to higher water uptake, and then consequently lower soil water content which led to higher gas diffusion and, thus, higher CH₄ oxidation. Methanotrophs also needs phosphorus for community structure growth.

While N is often the primary limiting nutrient in temperate and boreal forests, phosphorus usually limits ecological processes in tropical and subtropical forests (Vitousek and Sanford, 1986; Cleveland et al., 2002; Vitousek et al., 2010). We studied the relationships between soil P availability and

CH₄ flux in an old-growth tropical forest located in southern China where elevated N deposition has been well documented (Mo et al., 2006; Fang et al., 2008, 2011). Earlier studies in this old-growth forest showed that no N retention occurred, but rather a net loss of 8–16 kg N ha⁻¹ yr⁻¹ from the soil was estimated in the old-growth forest. In total, up to 60 kg N ha⁻¹ yr⁻¹ was leached from the old-growth forest, indicating that this forest was completely N saturated (Fang et al., 2008, 2011) from the chronic elevated N deposition in southern China. This interpretation is also supported by the results on litterfall production, which revealed no significant effects of N additions on total litterfall production in the old-growth forest (Mo et al., 2008). Studying forest CH₄ uptake and its relationship with soil available P and elevated N deposition is very important for evaluating the contribution of tropical forests to global climate change. Due to limited research in tropical forests, it is not clear how P availability would affect soil CH₄ uptake, and how P addition may interact with N deposition. This 33 months study experimentally tested the effects of P and N availabilities on soil CH₄ uptake. We hypothesized that: (1) N addition would inhibit soil CH₄ uptake as we found in a previous study in the same forest (Zhang et al., 2008); (2) P addition would increase soil CH₄ uptake due to that P addition would stimulate plant root growth and lead to higher water uptake, and then consequently lower WFPS which lead to higher gas diffusion and, thus, higher CH₄ uptake; (3) NP addition (interactive effect of N and P) would have less effect on soil CH₄ uptake comparing with N or P addition alone. This means that P addition would mitigate the inhibitive effect of N addition on soil CH₄ uptake.

2 Materials and methods

2.1 Site description

This study was conducted in the 1200 ha Dinghushan Biosphere Reserve (DHSBR), which is located in the middle of Guangdong Province, southern China (112°10' E, 23°10' N). There is an old-growth evergreen broadleaf forest (mature forest) in this reserve. The old-growth forest has been well protected from human activity for more than 400 years (Wang et al., 1982; Mo et al., 2003; Tang et al., 2006). The average annual precipitation of 1927 mm in the reserve has a distinct seasonal pattern, with 75 % falling from March to August and only 6 % falling from December to February (Huang and Fan, 1982). The mean annual temperature is 21 °C with the January mean temperature of 12.6 °C and July mean temperature of 28.0 °C (Huang and Fan, 1982). Annual mean relative humidity is 80 % (Huang and Fan, 1982). The wet N deposition was 36–38 kg N ha⁻¹ in the 1990s (Zhou and Yan, 2001). Precipitation N deposition in this region was 34.1 kg N ha⁻¹ yr⁻¹, with roughly 1.5:1 NH₄⁺ to NO₃⁻ ratio (Fang et al., 2011). Soil in the reserve is oxisols from

shale formation (Wu et al., 1982). The soil depth in the old-growth forest is more than 60 cm to the top of the C horizon (Mo et al., 2003). The forest in this experiment is situated on mountain slopes about 30°–35°.

2.2 Experimental treatment

Four treatments were established (each with five replicates) in 2007: control, N-addition (150 kg N ha⁻¹ yr⁻¹), P-addition (150 kg P ha⁻¹ yr⁻¹) and NP-addition (150 kg N ha⁻¹ yr⁻¹ plus 150 kg P ha⁻¹ yr⁻¹). A total of 20 plots of 5 m × 5 m were established and each plot was surrounded by a 5-m-wide buffer strip. Plots size and fertilizer level were referenced to the experiment in Costa Rica by Cleveland and Townsend (2006). Field plots and treatments were laid out randomly. NH₄NO₃ and NaH₂PO₄ solutions were sprayed once every other month to the forest floor with a backpack sprayer starting from February 2007 and continued through October 2009. Fertilizer was weighed and mixed with 5 l of water for each plot. Each control plot received 5 l of water without fertilizer.

2.3 Field sampling and measurements

CH₄, CO₂ and nitrous oxide (N₂O) flux were measured from January 2007 before the first fertilizer application. Static collars were installed in each plot in November 2006, two months before the gas sampling. Gas fluxes were monitored once every month using the static chamber and a gas chromatograph (Agilent 4890D). The static chamber was a 25-cm-diameter by 16-cm-tall PVC pipe permanently anchored 8 cm into the soil. During gas collection, a 30-cm-tall removable cover chamber was attached tightly to the anchor ring with a rubber band. Gas samples were collected from each chamber from 09:00–10:00 LT. Diurnal studies in the adjacent forests found that greenhouse gas fluxes measured during the mid-morning (09:00–10:00 LT) were closer to the daily mean (Tang et al., 2006). The GHG concentrations remained linear for up to 100 min after the chamber was closed in our study. Gas samples were taken with a 60 ml plastic syringe at 0 and 30 min after the chamber closure. Before each sampling, syringes were flushed three times with chamber gas to mix the headspace. Laboratory tests showed that chambers and syringes were inert to N₂O, CO₂ and CH₄ (Stuedler et al., 1989; Bowden et al., 1990). Gas samples were analysed within 12 h in a gas chromatograph (Agilent 4890D) equipped with a flame ionization detector (FID) for CH₄ and CO₂, and an electron capture detector (ECD) for N₂O. CO₂ was transformed into CH₄ via (Ni)H₂ before the FID analysis. Calibration gases (CH₄ at 1.87 ppm, CO₂ at 418 ppm, N₂O at 0.321 ppm, bottle's No. 070811) were obtained from the Institute of Atmospheric Physics, Chinese Academy of Sciences. In this paper, we report only data of soil CH₄ in the old-growth forest.

The calculation of GHG flux followed that described in Zhang et al. (2008), based on a linear regression of chamber gas concentration versus time (IAEA, 1992; Holland et al., 1999). Atmospheric pressure was measured at the sampling site using an air pressure gauge (Model THOMMEN 2000, Switzerland). Air temperature (enclosure), soil temperature (at 5 cm depth) and moisture (0–10 cm depth) were measured during each sampling. Soil moisture content was detected using a TDR-probe (Time Domain Reflectometry, Model Top TZS-I, China). Soil moisture (0–10 cm depth) values were converted to WFPS (Water Filled Pore Space) according to the following formula:

$$\text{WFPS (\%)} = \text{Vol (\%)} / (1 - \text{SBD}(\text{g cm}^{-3}) / 2.65(\text{g cm}^{-3})) \quad (1)$$

where SBD is soil bulk density, Vol is volumetric water moisture and 2.65 is the density of quartz.

Soil samples were collected in February 2007 (before the first fertilizer application) and February 2009 (after two years of fertilization) for the physical and chemical properties. Five soil cores (2.5 cm inner diameter) were collected randomly from each of the 20 plots at 0–10 cm soil depths and combined to one composite sample. The litter layer was carefully removed before the soil sampling. The pH of the soil sample was measured in a 1:2.5 soil/water suspension. Total N concentration was determined by the micro-Kjeldahl digestion followed by the analysis of ammonium on a Wescan ammonia analyser, while total P concentration was analysed colorimetrically after acidified ammonium persulfate digestion (Anderson and Ingram, 1989). Available P was extracted with 0.03 M ammonium fluoride and 0.025 M hydrochloric acid and analysed colorimetrically (Anderson and Ingram, 1989).

2.4 Statistical analysis

Repeated measures of Analysis of Variance (ANOVA, PROC MIXED with AR(1) from SAS – SAS Institute Inc., Cary NC, USA) was used to examine the effect of fertilizer treatments on soil GHG fluxes from February 2007 to October 2009. Two-way ANOVA (PROC GLM from SAS) was used to examine the effects of N and P addition. One-way ANOVA was used to examine the difference in soil pH, NH₄⁺, NO₃⁻ and available P among treatments. Linear regression analysis was performed by Origin 8.0 (OriginLab Corporation, Northampton, MA USA) to examine the relationship between CH₄ fluxes and soil WFPS contents and soil temperature. Out of 680 observations, three were identified as outliers, which were probably caused by chamber leaks, abnormally high WFPS, and other unknown factors, and were removed from the data analyses. Statistical significant differences were set at *p* values <0.05 unless otherwise stated. Mean values ± 1 standard error are given in the text.

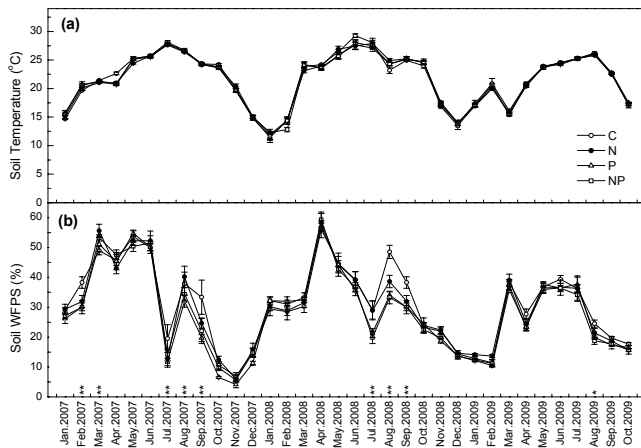


Fig. 1. Soil temperature at 5cm depth (a) and soil WFPS (b) during the study period. Error bars represent standard error of means (N=5). Asterisk (*) and double asterisk (**) indicates significant differences between control and at least one of the experiment treatments at $p < 0.1$ and $p < 0.05$, respectively.

3 Results

3.1 Soil temperature and WFPS

Soil temperature (at 5 cm depth) followed the air temperature in all plots, with temperatures increased from spring to summer and decreased from fall to winter (Fig. 1a). There was no treatment effect on soil temperature during the study period. Soil WFPS (0–10 cm depth) rose following the increased precipitation from dry winters to wet springs but decreased in summer, possibly due to plant uptake and higher evaporation, despite the high amount of precipitation in summer (Fig. 1b). Repeated measurement ANOVA showed that soil WFPS was significantly lower in the P-addition and NP-addition plots ($p = 0.011$ and $p = 0.018$, respectively) than in the control plots. However, there was no treatment effect in the N-addition plots ($p = 0.165$).

3.2 Other soil properties

Nitrogen and P treatments changed soil nutrient conditions (Fig. 2). Soil pH increased significantly in the P-addition plots ($p = 0.021$) and was a little higher in the NP-addition plots ($p = 0.062$), while no change in the N-addition plots ($p = 0.933$) (Fig. 2a) compared with the control plots. After 24 months of the treatment, a six-fold of soil available P was observed in P-addition plots ($p = 0.0003$) and four-fold in NP-addition plots ($p = 0.012$), compared to the controls (Fig. 2c). Furthermore, total P was significantly lower in N-addition plots compared to control plots ($p = 0.001$). This pattern is the same as pre-treatment (Fig. 2d). Soil NH_4^+ concentrations did not show any significant change in any treat-

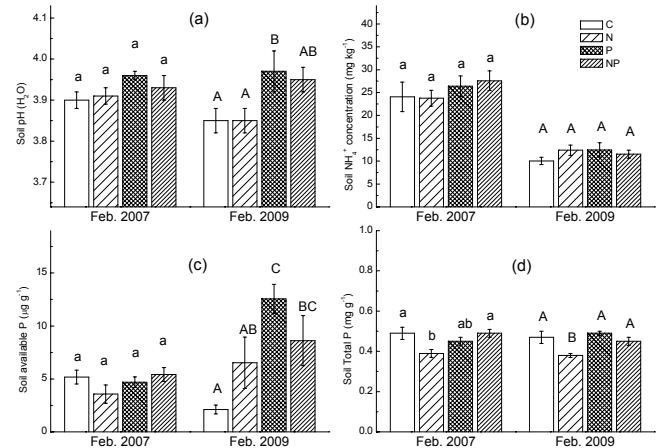


Fig. 2. Soil pH (a), ammonia nitrogen (b), available P (c) and total P (d) at 5 cm depth in February 2007 and February 2009. Error bars represent standard error of means (N=5). Different letters denote significant difference ($p < 0.05$) between treatments by the Repeated Measurement ANOVA.

ment plots (N-addition, P-addition and NP-addition) compared to the controls ($p = 0.163$, 0.152 and 0.368 , respectively) (Fig. 2b).

3.3 Soil CH_4 fluxes

Pre-treatment gas measurement (January 2007) showed no difference in different treatment plots compared to control plots ($p = 0.807$, 0.559 and 0.961 , respectively, in N, P, NP treatment plots). After fertilization treatment, repeated measurement ANOVA showed that P addition significantly increased soil CH_4 uptake while N addition significantly decreased CH_4 uptake (Fig. 3). The mean soil CH_4 uptake rate was $31.2 \pm 1.1 \mu\text{g CH}_4\text{-C m}^{-2} \text{h}^{-1}$ in the control plots during the 33 months study period. Soil CH_4 uptake in the P-addition plots was significantly higher (mean CH_4 uptake rate was $38.8 \pm 1.3 \mu\text{g CH}_4\text{-C m}^{-2} \text{h}^{-1}$, $p = 0.0068$). On the contrary, N-addition significantly inhibited soil CH_4 uptake (mean CH_4 uptake rate was $23.6 \pm 0.9 \mu\text{g CH}_4\text{-C m}^{-2} \text{h}^{-1}$, $p = 0.007$). No significant difference was observed between NP-addition plots ($33.6 \pm 1.0 \mu\text{g CH}_4\text{-C m}^{-2} \text{h}^{-1}$) and the controls ($p = 0.177$).

CH_4 uptake was higher in summer and fall (when the soil was low in water content) and was lower in spring (when the soil was wet) and winter (when the soil was cold). Two-way ANOVA showed a significant positive P effect on CH_4 uptake in the summers of 2007, 2008 and 2009, fall 2007, and the springs of 2008 and 2009 (Fig. 4), and a negative N effect in the springs of 2008 and 2009, and summer 2009 (Fig. 4). In summer 2007, CH_4 uptake in the P-addition and NP-addition plots was 26.6% and 27.3% higher than in the control plots ($p = 0.049$ and 0.051 , respectively). CH_4 uptake in the N-addition plots was reduced by 30.3% in fall

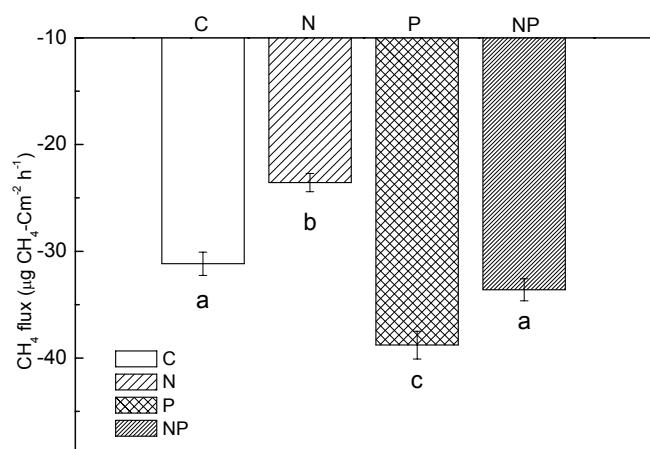


Fig. 3. Comparisons of mean soil CH_4 fluxes between treatments from 2007 to 2009 after N and P additions. Bars indicate ± 1 SE, $N=5$. Different letters denote significant difference ($p < 0.05$) between treatments by the Repeated Measurement ANOVA.

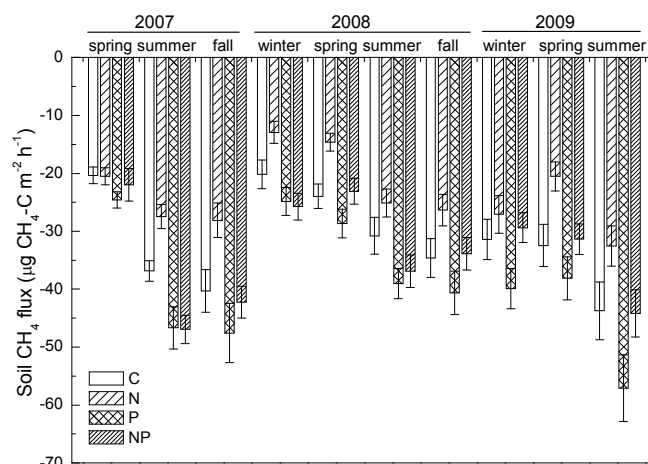


Fig. 4. Seasonal pattern of soil CH_4 uptake (April 2007 to September 2009). Bars indicate ± 1 SE, $N=5$. Spring from April to June, summer from July to September, fall from October to December and winter from January to March.

2007 ($p = 0.052$), 39.1% in spring 2008 ($p = 0.034$) and 36.7% in spring 2009 ($p = 0.027$). In addition, CH_4 uptake was increased by 30.6% in the P-addition plots in summer 2009 ($p = 0.038$).

CH_4 fluxes and soil WFPS were positively correlated in the control plots and N and P addition plots (Fig. 5). Under similar WFPS conditions, CH_4 uptake was the highest in P-addition plots and lowest in N-addition plots. CH_4 flux was not correlated to soil temperature.

4 Discussion

4.1 Compare with other tropical forests

The annual CH_4 uptake rates in our old-growth tropical forest site in southern China ranged from 2.3 to 3.4 $\text{kg CH}_4\text{-C ha}^{-1}\text{ yr}^{-1}$ (11 months in 2007, 12 months in 2008, 10 months in 2009), and were higher in the dry year (2007) than in the wet year (2008). The CH_4 fluxes, quantified in this study, are similar to the previous reported CH_4 fluxes measured at an adjacent forest (Tang et al., 2006; Zhang et al., 2008) and other parts of tropical Southwest China (Werner et al., 2006; Yan et al., 2008; Wang et al., 2010). The rates of CH_4 uptake in other tropical forests are also similar, ranging from 0.8 to 4.73 $\text{kg CH}_4\text{-C ha}^{-1}\text{ yr}^{-1}$ (Steudler et al., 1991; Kiese et al., 2003; Davidson and Nepstad, 2004; Davidson et al., 2008).

4.2 Effects of soil temperature and pH

Temperature variation has a minor impact on soil CH_4 uptake in our studied forest, which is consistent with previous results from the adjacent forest (Tang et al., 2006). In temperate or boreal regions, in contrast, soil CH_4 oxidation was generally positively correlated with soil temperature (Crill, 1991; King, 1997). Castro et al. (1995) observed that methanotrophy was affected by soil temperature between -5 and 10°C , but in our site, almost all of soil temperatures were between 10 and 30°C (Fig. 1a). The higher pH after P addition might contribute to the increase in CH_4 uptake rate in this forest. Most of the soils oxidised CH_4 over a pH range of $3\sim 7.5$ supporting in situ observations (Born et al., 1990). Bacteria extracted from boreal acidic forest soils had their highest CH_4 oxidation activity at pH 5.8 (Amaral et al., 1998). Soil pH in this acidic old-growth forest almost below 4 or even lower. Bradford et al. (2001b) observed that the drop in pH in a beech forest in the UK contributed to a decrease in soil CH_4 uptake. However, in some acidic peat soils, the rates of methane uptake were only slightly influenced by pH at values between 4.0 and 6.0 and decreased sharply at values below and above this range, although oxidation still occurred below pH 4.0 (Hanson and Hanson, 1996). Therefore, P addition increased soil pH in our study which may lead to increased methane oxidation.

4.3 Effects of soil WFPS

Soil CH_4 uptake was negatively correlated with the soil WFPS (a better measurement of soil moisture) in this study (Fig. 5), which was consistent with previous publications (Born et al., 1990; Castro et al., 1995; Kiese et al., 2003). Steinkamp et al. (2001) reported similarly that soil moisture was the dominant factor controlling CH_4 uptake when soil temperature was $>10^\circ\text{C}$. Methanotrophs use CH_4 as only a C and energy source and oxygen availability is the main factor limiting their activity (Le Mer and Roger, 2001). Lower

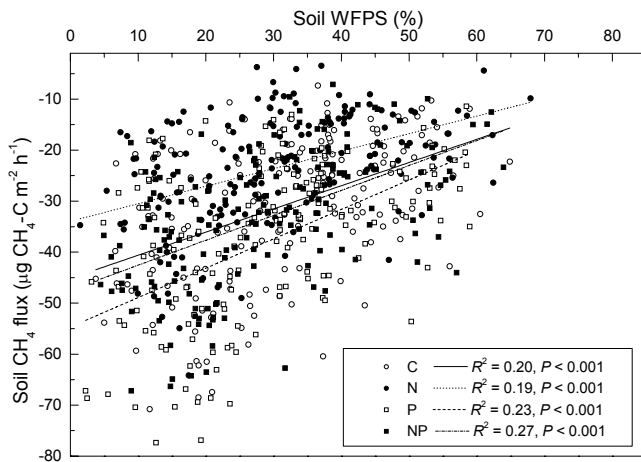


Fig. 5. Correlations between CH_4 flux and soil WFPS under different fertilization treatments. Under similar soil WFPS conditions, CH_4 uptake was higher in P-addition plots and lower in N-addition plots.

WFPS after P addition in our study might lead to higher soil aeration and more available oxygen and atmospheric methane to methanotrophs.

4.4 Effects of P and N fertilization

We found P addition significantly increased CH_4 uptake, N addition decreased CH_4 uptake, and P addition mitigated the negative N effect in this 33 months field experiment, as hypothesized. We believe this is the first experimental testing of N and P limitation on soil CH_4 flux in tropical and subtropical forests. The treatment effects were particularly strong in the summers and falls, when soil uptakes of CH_4 were the highest (Fig. 4). Phosphorus fertilization was often conducted in agricultural ecosystems. On planted rice soils, phosphorus addition significantly decreased CH_4 emission (Lu et al., 1999) probably by increasing methanotrophic potential (Joulian et al., 1998). We infer that this response of CH_4 uptake to the experimental P additions could be resulted from increased soil diffusion due to plant root growth. In our study, we found that soil WFPS was decreased after phosphorus addition, particularly in the summers (growing season) with a higher methane oxidation. This decreasing of soil WFPS was likely due to increased plant water consumption after P addition. Since phosphorus is a common limiting nutrient in tropical forests (Herbert et al., 2003; Wardle et al., 2004), low phosphorus availability due to strong sorption is often the main constraint on plant growth on highly weathered tropical soils (Vitousek, 1984). Ostertag (2001) also observed P-fertilizer leads to a greater root turnover in 4.1 million year-old forests in Hawaii. In these forests, two years fertilization of $150 \text{ kg ha}^{-1} \text{ yr}^{-1}$ did not increase total soil phosphorus probably due to serious plant uptake under high N deposition. Phosphorous deficiency in plant growth

in this old-growth forest is also partially supported by the results on litterfall study, which showed that annual total litterfall fluxes were significantly increased after P addition (Liu et al., 2011). Thus, P addition might stimulate plant roots sorption of phosphorus and water uptake which lead to lower soil WFPS in this forest. Higher water uptake by plant roots would subsequently result in higher soil diffusion. Therefore, higher diffusion for oxygen and atmospheric methane would increase methanotrophy activities. In addition, since phosphorus is a common limiting nutrient in tropical forests (Herbert et al., 2003; Wardle et al., 2004), microbial activity could be strongly P limited (Cleveland et al., 2002). Increased soil CH_4 uptake in this study may be due to stimulated methanotrophy activities after increased P availability in soil. In a companion study, we found that microbial biomass increased significantly by P addition (Liu et al., 2011). However, this could be a minor contribution to the increased CH_4 uptake due to methane oxidizing bacteria representing only a very small portion of the total soil microbial biomass. On the other hand, phosphorus fertilizer may increase the mineralization of organic P (Ofori-Frimpong and Rowell, 1999) and reduce Al^{3+} toxicity. Increased soil P levels might stabilize soil aluminum and iron through geochemical reaction of adsorption (Frossard et al., 1995). This coupling effect decreased aluminum toxicity to methanotrophs (Namba and King, 2000). However, in our experiment, we found no treatment effect on soil exchangeable Al^{3+} (data was not shown here).

The inhibitive effect of N input on CH_4 uptake has been reported extensively. Zhang et al. (2008) suggested that the response of forest soil CH_4 uptake on N fertilization possibly depends on soil N status. They compared the CH_4 uptake under various N additions in three tropical forests with very different N conditions and found that N addition in N-saturated old-growth forest significantly decreased soil CH_4 uptake while in N-limited young forests N addition had a limited effect. Singh et al. (1997) observed in a natural gradient of tropical forests, sites with higher soil mineral nitrogen concentrations had lower CH_4 uptake. In a Puerto Rican wet forest, Steudler et al. (1991) found that maximum reduction in CH_4 uptake coincided with the highest soil NH_4^+ , suggesting that nitrogen- CH_4 linkage observed in temperate forests (Steudler et al., 1989) may also function in tropical forests. NH_4^+ can reduce the growth rate of many methanotrophs by inhibiting CH_4 oxidation (Whittenbury et al., 1970). Laboratory studies indicate that oxidation of CH_4 by a variety of methanotrophs is competitively inhibited by nitrogen (Ferenci et al., 1975). In soils, the inhibition of CH_4 oxidation by ammonium is attributed to a competition at the level of the methane mono-oxygenase, a transfer of the methanotrophy activity towards nitrification (Castro et al., 1994) and the toxicity of NO_2 produced. Schnell and King (1994) observed that nitrite, the end product of methanotrophic ammonia oxidation, was a more effective inhibitor of CH_4 oxidation than ammonium. This inhibition becomes

irreversible and can only be released at CH₄ concentrations higher than 100 ppm (King and Schnell, 1994). In this old-growth forest, we observed that atmospheric CH₄ concentrations was around 2 ppm and even in the soil can not come up to 100 ppm. We suggest that the inhibition of CH₄ uptake by ammonium has occurred and persisted. However, we did not observe soil NH₄⁺ concentration change after treatments in this study, which might be due to the rapid volatile and leaching of the fertilization before our soil sampling.

5 Conclusions

Nitrogen deposition would weaken the function of upland forest soils as CH₄ sinks by its inhibitive effect on soil CH₄ uptake and by decreasing availability of phosphorus due to continuous nitrogen deposition and phosphorus limitation in the tropical forests of southern China and other parts of the world. Galloway et al. (1994) forecasted a large increase in nitrogen deposition from fossil-fuel, growing population and agricultural activity for year 2020 in those regions. However, P addition could reverse such trends by stimulating forest soil CH₄ uptake. In this 33 months field experiment, we showed that increased P availability might mitigate the inhibitive effect of N addition on soil CH₄ uptake in this N-saturated old-growth tropical forest. While average annual consumption of CH₄ was enhanced after P addition and decreased after N addition, there was no difference between the NP-addition plots and the controls (i.e., the negative effect of N addition on CH₄ uptake was reduced by P addition). As far as we know, our study is among the first field study experimentally testing the responses of soil CH₄ uptake to N and P additions in tropical forests. Results from our study suggest that phosphorus fertilization in tropical forests could be one of the future choices for mitigating the inhibitive effect of N deposition on soil CH₄ uptake.

Supplementary material related to this article is available online at:

<http://www.biogeosciences.net/8/2805/2011/bg-8-2805-2011-supplement.pdf>.

Acknowledgements. We would also like to thank Sandra Brown, Per Gundersen, Guirui Yu, Linghao Li and Wei Zhang for their constructive suggestions to this paper. Timothy Scott from the Binghamton University provided valuable editorial comments. This study was founded by the National Key Basic Research 973 Programme (2010CB833502), National Natural Science Foundation of China (Nos. 30970521, 40730102) and Natural Science Foundation of Guangdong Province, China (Grant 8351065005000001).

Edited by: X. Wang

References

- Amaral, J. A., Ren, T., and Knowles, R.: Atmospheric Methane Consumption by Forest Soils and Extracted Bacteria at Different pH Values, *Appl. Environ. Microbiol.*, 64, 2397–2402, 1998.
- Anderson, J. and Ingram, J.: *Tropical soil biology and fertility: A handbook of methods*. CAB International, Commonwealth Agricultural Bureaux (CAB) International, Wallingford, UK, 221 pp., 1989.
- Bodelier, P. L. E. and Laanbroek, H. J.: Nitrogen as a regulatory factor of methane oxidation in soils and sediments, *FEMS Microbiol. Ecol.*, 47, 265–277, 2004.
- Born, M., Dörr, H., and Levin, I.: Methane consumption in aerated soils of the temperate zone, *Tellus B*, 42, 2–8, 1990.
- Bowden, R., Steudler, P., Melillo, J., and Aber, J.: Annual nitrous oxide fluxes from temperate forest soils in the northeastern United States, *J. Geophys. Res.-Atmos.*, 95, 13997–14005, 1990.
- Börjesson, G. and Nohrstedt, H.: Fast recovery of atmospheric methane consumption in a Swedish forest soil after single-shot N-fertilization, *Forest Ecol. Manag.*, 134, 83–88, 2000.
- Bradford, M. A., Ineson, P., Wookey, P. A., and Lappin-Scott, H. M.: The effects of acid nitrogen and acid sulphur deposition on CH₄ oxidation in a forest soil: a laboratory study, *Soil Biol. Biochem.*, 33, 1695–1702, 2001a.
- Bradford, M. A., Wookey, P. A., Ineson, P., and Lappin-Scott, H. M.: Controlling factors and effects of chronic nitrogen and sulphur deposition on methane oxidation in a temperate forest soil, *Soil Biol. Biochem.*, 33, 93–102, 2001b.
- Butterbach-Bahl, K. and Papen, H.: Four years continuous record of CH₄-exchange between the atmosphere and untreated and limed soil of a N-saturated spruce and beech forest ecosystem in Germany, *Plant Soil*, 240, 77–90, 2002.
- Castro, M., Peterjohn, W., Melillo, J., Steudler, P., Gholz, H., and Lewis, D.: Effects of nitrogen fertilization on the fluxes of N₂O, CH₄, and CO₂ from soils in a Florida slash pine plantation, *Canadian journal of forest research(Print)*, 24, 9–13, 1994.
- Castro, M. S., Steudler, P. A., Melillo, J. M., Aber, J. D., and Bowden, R. D.: Factors controlling atmospheric methane consumption by temperate forest soils, *Global Biogeochem. Cy.*, 9, 1–10, 1995.
- Chan, A. S. K., Steudler, P. A., Bowden, R. D., Gulledge, J., and Cavanaugh, C. M.: Consequences of nitrogen fertilization on soil methane consumption in a productive temperate deciduous forest, *Biol. Fert. Soils*, 41, 182–189, 2005.
- Cleveland, C. C., Townsend, A. R., and Schmidt, S. K.: Phosphorus limitation of microbial processes in moist tropical forests: Evidence from short-term laboratory incubations and field studies, *Ecosystems*, 5, 680–691, 2002.
- Cleveland, C. C. and Townsend, A. R.: Nutrient additions to a tropical rain forest drive substantial soil carbon dioxide losses to the atmosphere, *P. Natl. Acad. Sci.*, 103, 10316–10321, 2006.
- Crill, P. M.: Seasonal patterns of methane uptake and carbon dioxide release by a temperate woodland soil, *Global Biogeochem. Cy.*, 5, 319–334, 1991.
- Davidson, E. A. and Nepstad, D. C.: Effects of an experimental drought on soil emissions of carbon dioxide, methane, nitrous oxide, and nitric oxide in a moist tropical forest, *Glob. Change Biol.*, 10, 718–730, 2004.
- Davidson, E. A., Nepstad, D. C., Ishida, F. Y., and Brando, P. M.: Effects of an experimental drought and recovery on soil emis-

- sions of carbon dioxide, methane, nitrous oxide, and nitric oxide in a moist tropical forest, *Glob. Change Biol.*, 14, 2582–2590, 2008.
- Fang, Y., Yoh, M., Koba, K., Zhu, W., Takebayashi, Y. U., Xiao, Y., Lei, C., Mo, J., Zhang, W., and Lu, X.: Nitrogen deposition and forest nitrogen cycling along an urban-rural transect in southern China, *Glob. Change Biol.*, 17, 872–885, 2011.
- Fang, Y. T., Gundersen, P., Mo, J. M., and Zhu, W. X.: Input and output of dissolved organic and inorganic nitrogen in subtropical forests of South China under high air pollution, *Biogeosciences*, 5, 339–352, doi:10.5194/bg-5-339-2008, 2008.
- Ferenci, T., Strom, T., and Quayle, J. R.: Oxidation of Carbon Monoxide and Methane by *Pseudomonas methanica*, *J. Gen. Microbiol.*, 91, 79–91, 1975.
- Frossard, E., Brossard, M., Hedley, M., and Metherell, A.: Reactions controlling the cycling of P in soils, *Scope-Scientific Committee on Problems of the Environment International Council of Scientific Unions*, 54, 107–138, 1995.
- Galloway, J. N., Levy, H., and Kashibhatla, P. S.: Year 2020: Consequences of population growth and development on deposition of oxidized nitrogen, *Ambio*, 23, 120–123, 1994.
- Hanson, R. S. and Hanson, T. E.: Methanotrophic bacteria, *Microbiol. Rev.*, 60, 439–471, 1996.
- Hein, R., Crutzen, P. J., and Heimann, M.: An inverse modeling approach to investigate the global atmospheric methane cycle, *Global Biogeochem. Cy.*, 11, 43–76, 1997.
- Herbert, D. A., Williams, M., and Rastetter, E. B.: A model analysis of N and P limitation on carbon accumulation in Amazonian secondary forest after alternate land-use abandonment, *Biogeochemistry*, 65, 121–150, 2003.
- Holland, E. A., Robertson, G. P., Greenberg, J., Groffman, P. M., Boone, R. D., and Gosz, J. R.: Soil CO₂, N₂O, and CH₄ exchange, in: *Standard Soil Methods for Long-term Ecological Research*, edited by: Robertson, G. P., Oxford University Press, USA, 185–201, 1999.
- Huang, Z. F. and Fan, Z. G.: The climate of Dinghushan (in Chinese with English abstract), *Tropical and Subtropical Forest Ecosystem*, 1, 11–16, 1982.
- IAEA: *Manual on Measurements of Methane and Nitrous Oxide Emission from Agriculture*, International Atomic Energy Agency, Vienna, Austria, 91 pp., 1992.
- IPCC: *Changes in atmospheric constituents and in radiative forcing*, in: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, edited by: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K. B., Tignor, M., and Miller, H. L., Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 130–234, 2007.
- Jamali, H., Livesley, S. J., Grover, S. P., Dawes, T. Z., Hutley, L. B., Cook, G. D., and Arndt, S. K.: The Importance of Termites to the CH₄ Balance of a Tropical Savanna Woodland of Northern Australia, *Ecosystems*, 14, 698–709, 2011.
- Joulian, C., Ollivier, B., Patel, B. K. C., and Roger, P. A.: Phenotypic and phylogenetic characterization of dominant culturable methanogens isolated from ricefield soils, *FEMS Microbiol. Ecol.*, 25, 135–145, 1998.
- Kiese, R., Hewett, B., Graham, A., and Butterbach-Bahl, K.: Seasonal variability of N₂O emissions and CH₄ uptake by tropical rainforest soils of Queensland, Australia, *Global Biogeochem. Cy.*, 17, 1043–1055, 2003.
- King, G.: Responses of atmospheric methane consumption by soils to global climate change, *Glob. Change Biol.*, 3, 351–362, 1997.
- King, G. M. and Schnell, S.: Effect of increasing atmospheric methane concentration on ammonium inhibition of soil methane consumption, *Nature*, 370, 282–284, 1994.
- Kolb, S.: The quest for atmospheric methane oxidizers in forest soils, *Environmental Microbiology Reports*, 1, 336–346, 2009.
- Le Mer, J. and Roger, P.: Production, oxidation, emission and consumption of methane by soils: A review, *Eur. J. Soil Biol.*, 37, 25–50, 2001.
- Lelieveld, J., Crutzen, P. J., and Dentener, F. J.: Changing concentration, lifetime and climate forcing of atmospheric methane, *Tellus B*, 50, 128–150, 1998.
- Liu, L. L. and Greaver, T. L.: A review of nitrogen enrichment effects on three biogenic GHGs: the CO₂ sink may be largely offset by stimulated N₂O and CH₄ emission, *Ecol. Lett.*, 12, 1103–1117, 2009.
- Liu, L., Gundersen, P., Zhang, T., and Mo, J. M.: Effects of phosphorus addition on soil microbial biomass and community composition in three forest types in tropical China, *Soil Biol. Biochem.*, doi:10.1016/j.soilbio.2011.08.017, 2011.
- Lu, Y., Wassmann, R., Neue, H. U., and Huang, C.: Impact of Phosphorus Supply on Root Exudation, Aerenchyma Formation and Methane Emission of Rice Plants, *Biogeochemistry*, 47, 203–218, 1999.
- Menyailo, O., Abraham, W., and Conrad, R.: Tree species affect atmospheric CH₄ oxidation without altering community composition of soil methanotrophs, *Soil Biol. Biochem.*, 42, 101–107, 2010.
- Mo, J. M., Brown, S., Peng, S. L., and Kong, G. H.: Nitrogen availability in disturbed, rehabilitated and mature forests of tropical China, *Forest Ecol. Manag.*, 175, 573–583, 2003.
- Mo, J. M., Brown, S., Xue, J. H., Fang, Y. T., and Li, Z. A.: Response of litter decomposition to simulated N deposition in disturbed, rehabilitated and mature forests in subtropical China, *Plant Soil*, 282, 135–151, 2006.
- Mo, J., Zhang, W., Zhu, W., Gundersen, P., Fang, Y., Li, D., and Wang, H.: Nitrogen addition reduces soil respiration in a mature tropical forest in southern China, *Glob. Change Biol.*, 14, 403–412, 2008.
- Mohanty, S. R., Bodelier, P. L. E., and Conrad, R.: Effect of temperature on composition of the methanotrophic community in rice field and forest soil, *FEMS Microbiol. Ecol.*, 62, 24–31, 2007.
- Nanba, K. and King, G. M.: Response of atmospheric methane consumption by Maine forest soils to exogenous aluminum salts, *Applied and Environmental Microbiology*, 66, 3674–3679, 2000.
- Ofori-Frimpong, K. and Rowell, D. L.: The decomposition of cocoa leaves and their effect on phosphorus dynamics in tropical soil, *Eur. J. Soil Biol.*, 50, 165–172, 1999.
- Ostertag, R.: Effects of nitrogen and phosphorus availability on fine-root dynamics in Hawaiian montane forests, *Ecology*, 82, 485–499, 2001.
- Schnell, S. and King, G. M.: Mechanistic Analysis of Ammonium Inhibition of Atmospheric Methane Consumption in Forest Soils, *Appl. Environ. Microbiol.*, 60, 3514–3521, 1994.
- Singh, J. S., Singh, S., Raghubanshi, A. S., Singh, S., Kashyap, A. K., and Reddy, V. S.: Effect of soil nitrogen, carbon and moisture

- on methane uptake by dry tropical forest soils, *Plant Soil*, 196, 115–121, 1997.
- Steinkamp, R., Butterbach-Bahl, K., and Papen, H.: Methane oxidation by soils of an N limited and N fertilized spruce forest in the Black Forest, Germany, *Soil Biol. Biochem.*, 33, 145–153, 2001.
- Stuedler, P. A., Bowden, R. D., Melillo, J. M., and Aber, J. D.: Influence of nitrogen fertilization on methane uptake in temperate forest soils, *Nature*, 341, 314–316, 1989.
- Stuedler, P. A., Melillo, J. M., Bowden, R. D., Castro, M. S., and Lugo, A. E.: The effects of natural and human disturbances on soil nitrogen dynamics and trace gas fluxes in a Puerto Rican wet forest, *Biotropica*, 23, 356–363, 1991.
- Tang, X. L., Liu, S. G., Zhou, G. Y., Zhang, D. Q., and Zhou, C. Y.: Soil-atmospheric exchange of CO₂, CH₄, and N₂O in three subtropical forest ecosystems in southern China, *Glob. Change Biol.*, 12, 546–560, 2006.
- Tate, K. R., Ross, D. J., Saggarr, S., Hedley, C. B., Dando, J., Singh, B. K., and Lambie, S. M.: Methane uptake in soils from *Pinus radiata* plantations, a reverting shrubland and adjacent pastures: Effects of land-use change, and soil texture, water and mineral nitrogen, *Soil Biol. Biochem.*, 39, 1437–1449, 2007.
- Teh, Y. A., Silver, W. L., and Conrad, M. E.: Oxygen effects on methane production and oxidation in humid tropical forest soils, *Glob. Change Biol.*, 11, 1283–1297, 2005.
- Templeton, A. S., Chu, K. H., Alvarez-Cohen, L., and Conrad, M. E.: Variable carbon isotope fractionation expressed by aerobic CH₄-oxidizing bacteria, *Geochim Cosmochim. Acta.*, 70, 1739–1752, 2006.
- Vitousek, P. M.: Litterfall, Nutrient Cycling, and Nutrient Limitation in Tropical Forests, *Ecology*, 65, 285–298, 1984.
- Vitousek, P. M. and Sanford, R. L.: Nutrient cycling in moist tropical forest, *Ann. Rev. Ecol. Syst.*, 17, 137–167, 1986.
- Vitousek, P. M., Porder, S., Houlton, B. Z., and Chadwick, O. A.: Terrestrial phosphorus limitation: mechanisms, implications, and nitrogen-phosphorus interactions, *Ecol. Appl.*, 20, 5–15, 2010.
- Wang, H., Liu, S. R., Mo, J. M., and Zhang, T.: Soil-atmosphere exchange of greenhouse gases in subtropical plantations of indigenous tree species, *Plant Soil*, 335, 213–227, 2010.
- Wang, Z. H., He, D. Q., Song, S. D., Chen, S. P., Chen, D. R., and Tu, M. Z.: The vegetation of Dinghushan Biosphere Reserve (in Chinese with English abstract), *Tropical and Subtropical Forest Ecosystem*, 1, 77–141, 1982.
- Wardle, D. A., Walker, L. R., and Bardgett, R. D.: Ecosystem properties and forest decline in contrasting long-term chronosequences, *Science*, 305, 509–513, 2004.
- Werner, C., Zheng, X. H., Tang, J. W., Xie, B. H., Liu, C. Y., Kiese, R., and Butterbach-Bahl, K.: N₂O, CH₄ and CO₂ emissions from seasonal tropical rainforests and a rubber plantation in southwest China, *Plant Soil*, 289, 335–353, 2006.
- Whittenbury, R., Phillips, K. C., and Wilkinson, J. F.: Enrichment, isolation and some properties of methane-utilizing bacteria, *J. Gen. Microbiol.*, 61, 205–218, 1970.
- WMO: The state of greenhouse gases in the atmosphere based on global observations through 2009, *WMO Greenhouse Gas Bulletin*, 6, 1–4, 2010.
- Wu, H. S., Deng, H. Z., and Chen, H. T.: Physico-geographical features of Dinghushan and their dynamic analyses (in Chinese with English abstract), *Tropical and Subtropical Forest Ecosystem*, 1, 1–10, 1982.
- Yan, Y. P., Sha, L. Q., Cao, M., Zheng, Z., Tang, J. W., Wang, Y. Y., Zhang, Y. P., Wang, R., Liu, G. R., Wang, Y. S., and Sun, Y.: Fluxes of CH₄ and N₂O from soil under a tropical seasonal rain forest in Xishuangbanna, southwest China, *J. Environ. Sci.-China*, 20, 207–215, 2008.
- Zhang, W., Mo, J. M., Zhou, G. Y., Gundersen, P., Fang, Y. T., Lu, X. K., Zhang, T., and Dong, S. F.: Methane uptake responses to nitrogen deposition in three tropical forests in southern China, *J. Geophys. Res.-Atmos.*, 113, D11116, doi:10.1029/2007jd009195, 2008.
- Zhou, G. Y. and Yan, J. H.: The influence of region atmospheric precipitation characteristics and its element inputs on the existence and development of Dinghushan forest ecosystems (in Chinese with English abstract), *Acta Ecologica Sinica*, 21, 2002–2012, 2001.