

Carbon and oxygen isotope analysis of leaf biomass reveals contrasting photosynthetic responses to elevated CO₂ near geologic vents in Yellowstone National Park

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Abstract. In this study we explore the use of natural CO₂ emissions in Yellowstone National Park (YNP) in Wyoming, USA to study responses of natural vegetation to elevated CO₂ levels. Radiocarbon (¹⁴C) analysis of leaf biomass from a conifer (*Pinus contortus*; lodgepole pine) and an invasive, non-native herb (*Linaria dalmatica*; Dalmation toadflax) was used to trace the inputs of vent CO₂ and quantify assimilation-weighted CO₂ concentrations experienced by individual plants near vents and in comparable locations with no geologic CO₂ exposure. The carbon and oxygen isotopic composition and nitrogen percent of leaf biomass from the same plants was used to investigate photosynthetic responses of these plants to naturally elevated atmospheric CO₂ concentrations. The coupled shifts in carbon and oxygen isotope values suggest that dalmation toadflax responded to elevated CO₂ exposure by increasing stomatal conductance with no change in photosynthetic capacity and lodgepole pine apparently responded by decreasing stomatal conductance and photosynthetic capacity. Lodgepole pine saplings exposed to elevated levels of CO₂ likewise had reduced leaf nitrogen concentrations compared to plants with no enhanced CO₂ exposure, further suggesting widespread and dominant conifer down-regulated photosynthetic capacity under elevated CO₂ levels near geologic vents.

1 Introduction

Effects of elevated CO₂ on plant photosynthesis and growth have been intensively studied because of the prominent role terrestrial vegetation plays in the global carbon cycle. Unresolved is whether and to what degree photosynthesis will be stimulated and how stomatal conductance and carboxylation and photochemical capacities are likely to react to globally increased CO₂ levels (Ainsworth and Rogers, 2007). Free-Air CO₂ Enrichment (FACE) is the preferred approach for studying effects of elevated CO₂ on natural ecosystems under realistic soil conditions and climate. However, cost constraints and technical difficulties have severely limited application of FACE technology across different ecosystems and climate. Further due to cost constraints most FACE experiments are short term providing only limited opportunities to study feedbacks on community structure and soil biogeochemistry. Geologic CO₂ emissions at the terrestrial surface offer an alternative low cost approach for studying elevated CO₂ effects on plant communities growing in natural environments (Bettaraini et al., 1995; Miglietta et al., 1998; Tognetti et al., 2000). Although natural CO₂ gradients established near geologic CO₂ vents can be confounded with other environmental factors, observations from such systems provide an opportunity to examine effects of long-term exposure to elevated CO₂ to compare against results from short-term Free-Air CO₂ Enrichment (FACE) experiments.

In this study we explore the use of natural CO₂ emissions in Yellowstone National Park (YNP) in Wyoming, USA to study responses of a native conifer (*Pinus contortus*; Lodgepole pine) and an invasive, non-native herb (*Linaria dalmatica*; Dalmation toadflax) to elevated CO₂ levels. There are



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numerous cold geologic features in YNP emitting large quantities of CO₂ at very high concentrations (>99%) with very minimal traces of H₂S and CH₄ and the native vegetation has been exposed to elevated levels of CO₂ at these sites for generations. The CO₂ emitted from geologic vents in YNP has no ¹⁴C, whereas background air has distinguishable and stable levels of ¹⁴CO₂ incorporated into plant biomass during photosynthesis. Hence, radiocarbon (¹⁴C) analysis of leaf biomass was used to trace the inputs of vent CO₂ and quantify assimilation-weighted CO₂ concentrations experienced by individual plants near vents and in comparable locations with no geologic CO₂ exposure.

Changes in plant photosynthesis associated with increased atmospheric CO₂ concentration can be detected in the ¹³C and ¹⁸O isotope composition of leaves (Farquhar et al., 1989; Ehleringer et al., 1993; Scheidegger et al., 2000; Barbour, 2007). The combined ¹³C and ¹⁸O isotopic composition of leaves can be used as long-term integrators of plant response to elevated CO₂ conditions (Grams et al., 2007). Increases in leaf carbon isotope discrimination can be a result of either (1) a proportional increase in stomatal conductance relative to photosynthetic CO₂ demand by chloroplasts or (2) a proportional decrease in photosynthetic CO₂ demand by chloroplasts relative to stomatal conductance (Scheidegger et al., 2000). Measurements of the carbon and oxygen isotopic composition of leaf organic matter can be used to investigate independent effects of photosynthetic capacity and stomatal conductance on the ratio of internal to ambient CO₂ composition (*c_i/c_a*) and hence Δ¹³C (Scheidegger et al., 2000; Barbour, 2007; Grams et al., 2007). The combined assessment of δ¹³C and δ¹⁸O and N content of collected leaf samples helped us to investigate photosynthetic responses of both plant species to naturally elevated atmospheric CO₂ concentrations.

2 Methods

2.1 Leaf sampling and determination of CO₂ exposure

Foliage samples were collected from lodgepole pine saplings at Ochre Springs and from adult plants of the invasive, non-native forb dalmation toadflax at Mammoth Upper Terraces in July 2006 (Fig. 1). Plants were sampled from near low-temperature CO₂ vents (“elevated” CO₂ sites) and from nearby sites where CO₂ exposure from vents was minimal (“control” CO₂ sites). Sampling locations were selected based on observed atmospheric CO₂ concentration values measured with a portable infra-red gas analyzer (IRGA; LI 6200, LiCor Inc., Lincoln NE, USA) and from site surveys (Tereck, 2004). Instantaneous CO₂ concentrations within plant canopies measured with the IRGA were elevated (400–600 μL/L) above background levels near the CO₂ vents and were near background levels (378 μL/L) at comparable locations distant from vents. The control CO₂ site at Ochre

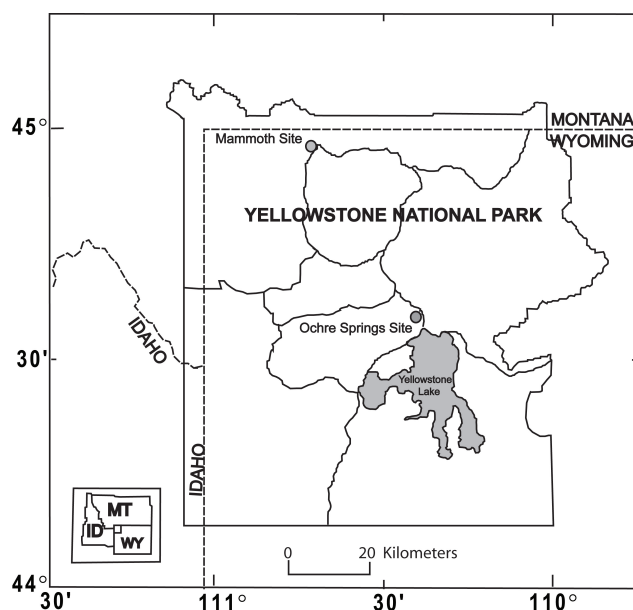


Fig. 1. Map of Yellowstone National Park showing the Ochre Spring and Mammoth Hot Springs sampling locations.

Springs (44°37'1" N; –110°26'12" W) was located approximately 1600 m from the elevated CO₂ site (44°36'45" N; –110°24'52" W). At Mammoth Upper Terraces the control site (44°57'42" N; –110°42'51" W) and elevated CO₂ site (44°57'45" N; –110°42'54" W) were at distance of approximately 200 m from each other. Soil temperature variations can potentially effect the photosynthesis and water relations in different plant communities (Schwarz et al., 1997; Dang and Cheng, 2004; Zhang and Dang, 2005). Therefore, soil temperatures were measured at all sampling sites and we noted that soil temperatures at 10 cm depth were similar (between 20 and 25°C at midday) at both control and elevated CO₂ sites.

Five individuals of lodgepole pine were haphazardly selected for isotope sampling at each of the control and elevated CO₂ sites in the Ochre Springs area. Isotopic composition of leaves might be effected by increased hydraulic resistance resulting from increased xylem path lengths in taller trees and longer branches (Warren and Adams, 2000) and the effective CO₂ exposure could also vary considerably with canopy height. To reduce this variability in this study needles were sampled (from the 2006 cohort) from branches of same length at the middle crown position from 0.5 to 1 m tall trees. The light environment of leaves can also potentially affect their carbon isotope signature (Lockheart et al., 1997; Grams et al., 2007). Therefore, we sampled needles only from tips of south facing branches of trees exposed to similar microhabitat conditions within the forest stand. Leaves of dalmation toadflax at Mammoth Upper Terraces were collected from 5 plants growing near a point CO₂ emission source and from 4 toadflax plants growing in a nearby site

where atmospheric CO₂ levels were similar to background levels. At both the elevated and control site leaves were sampled from 4 to 5 plants growing at a distance of 3–4 m from each other along a 15 m transect. Leaves (10–15) were collected from 4 different stems on each plant avoiding the small leaves subtending flowers.

Quantitative exposure to CO₂ from the geologic emissions integrated over the growth period of leaf development and maturation is difficult to assess solely from short-term IRGA measurements of air in the plant canopy. Therefore, we used the ¹⁴C content of leaves to estimate the assimilation-weighted CO₂ concentration exposure. The CO₂ at our sampling sites can only come from 2 sources i.e. atmosphere or the vent and therefore the sum of fractions of CO₂ fixed by plant coming from the vent and normal atmosphere should be equal to unity. Using mass balance approach, the measured ¹⁴C abundance in plant material (M_p) will be equal to the sum of fractions coming from vent and atmosphere weighted by the abundance of ¹⁴C in each of these gases (van Gardingen et al., 1995). CO₂ emitted from the vents originates from dissolution of carbonate sediments and therefore is highly depleted in ¹⁴C (so called “dead CO₂”) and ¹⁴C contribution from vent can be taken as 0 (Bettarini et al., 1995; van Gardingen et al., 1995). A simple mass-balance approach was then used to calculate the average CO₂ concentration surrounding individual lodgepole pine and dalmation toadflax plants:

$$[\text{CO}_2]_p = [\text{CO}_2]_b(M_b/M_p) \quad (\text{R1})$$

where $[\text{CO}_2]_b$ is the mean CO₂ concentration in the normal background atmosphere (taken to be 380 ppm). M_b and M_p are the ¹⁴C abundances (reported as fraction of modern standard) in leaves growing at ambient CO₂ concentrations and higher CO₂ concentrations respectively.

Carbon isotope discrimination was determined from $\delta^{13}\text{C}$ of leaf biomass ($\delta^{13}\text{C}_l$) and $\delta^{13}\text{C}$ of air surrounding the plant ($\delta^{13}\text{C}_p$) from (Farquhar et al., 1989):

$$\Delta^{13}\text{C} = (\delta^{13}\text{C}_p - \delta^{13}\text{C}_l)/(1 + \delta^{13}\text{C}_l/1000) \quad (\text{R2})$$

The $\delta^{13}\text{C}$ of CO₂ in air surrounding the plant ($\delta^{13}\text{C}_p$) will represent a mixture of $\delta^{13}\text{C}$ of CO₂ from the vent ($\delta^{13}\text{C}_v$) and background air ($\delta^{13}\text{C}_b$). CO₂ emitted from geologic vents at Mammoth Upper Terraces and Ochre Spring sites has $\delta^{13}\text{C}$ values of −3.9 and −3.2‰, respectively (Werner and Brantley, 2003). The $\delta^{13}\text{C}$ of background atmospheric CO₂ was assumed to be −8‰ (Keeling et al., 2001). The assimilation-weighted $\delta^{13}\text{C}$ value of CO₂ in the air surrounding individual plants was calculated using mass-balance:

$$\delta^{13}\text{C}_p = \delta^{13}\text{C}_b + f(\delta^{13}\text{C}_v - \delta^{13}\text{C}_b) \quad (\text{R3})$$

where f is the fraction of CO₂ contribution from the vent. The fraction of CO₂ contributed by vent is calculated using mass balance approach:

$$f = (\Delta^{14}\text{C}_l - \Delta^{14}\text{C}_b)/(\Delta^{14}\text{C}_v - \Delta^{14}\text{C}_b) \quad (\text{R4})$$

where $\Delta^{14}\text{C}$ is the relative deviation of ¹⁴C activity of the sample from the oxalic Acid I standard in per mil values (after accounting for ¹⁴C fractionation with a $\delta^{13}\text{C}$ correction). $\Delta^{14}\text{C}_l$ is the measured value of leaf samples, $\Delta^{14}\text{C}_b$ is the measured value for 2006 atmosphere at Pt. Barrow Alaska (50‰) and $\Delta^{14}\text{C}_v$ is the assumed value for CO₂ from vent (−1000‰).

The calculated carbon isotope discrimination was then used to estimate the influence of elevated CO₂ concentration on the ratio of leaf intercellular to ambient CO₂ concentration (c_i/c_a) using the simplified model of Farquhar et al. (1989) as:

$$c_i/c_a = (\Delta^{13}\text{C} - a)/(b - a) \quad (\text{R5})$$

where a is the carbon isotope discrimination associated with CO₂ diffusion through air (−4.4‰), and b is the net discrimination associated with CO₂ fixation in C3 plants (−27‰).

2.2 Leaf isotopic and N content analyses

Whole leaf samples of dalmation toadflax and lodgepole pine needles from the 2006 growth year were dried and ground in a mill to a consistent particle size. Stable isotope analysis was carried out at the University of Wyoming Stable Isotope Facility (<http://www.uwyo.edu/sif/>). The $\delta^{13}\text{C}$ and N‰ values were determined by continuous flow EA-IRMS using a Costech 4010 element analyzer online with a Finnigan Delta plus XP isotope ratio mass spectrometer. The $\delta^{18}\text{O}$ composition of bulk leaf material was measured by online pyrolysis to CO using a Finnigan TC/EA coupled to the Finnigan Delta Plus XP isotope ratio mass spectrometer. Measurement precision was assessed by replicate analysis of samples and internal lab standards and was better than ±0.1‰ for $\delta^{13}\text{C}$ and ±0.25‰ for $\delta^{18}\text{O}$. The $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ values are reported in per mil (‰) relative to V-PDB and V-SMOW, respectively. Radiocarbon analysis was performed at the W.M. Keck AMS Laboratory at the University of California, Irvine. Radiocarbon concentrations are reported as fractions of the Modern Standard and $\Delta^{14}\text{C}$ is reported following conventions of Stuiver and Polach (1977).

3 Results and discussion

The assimilation-weighted CO₂ exposure of a leaf calculated from radiocarbon values of leaves Eq. (1) ranged from 379–381 (control site) to 401–607 (elevated site) ppm in dalmation toadflax from the Mammoth Upper Terraces site and from 378–383 (control site) to 419–482 ppm (elevated site) in lodgepole pine at the Ochre Springs site. Changes in leaf $\delta^{13}\text{C}$ values in response to variations in environmental conditions integrate changes in the relative demand of CO₂ by chloroplasts during photosynthesis and in the supply of CO₂ associated with stomatal conductance in plants (Farquhar et al., 1989; Ehleringer et al., 1993). We

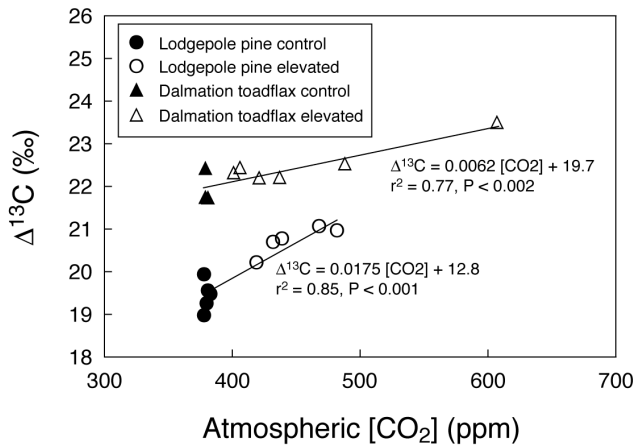


Fig. 2. Correlation between carbon isotope discrimination ($\Delta^{13}\text{C}$) and average flux-weighted CO₂ in the leaves of dalmation toadflax (open and filled triangles) and lodgepole pine (open and filled circles) were exposed to. The flux-weighted CO₂ exposure was calculated from the radiocarbon or ¹⁴C content of leaves using Eq. (1).

found that $\Delta^{13}\text{C}$ was significantly positively correlated with assimilation-weighted CO₂ exposure levels (Fig. 2) for both dalmation toadflax and lodgepole pine (dalmation toadflax, $r^2=0.77$, $P<0.002$; lodgepole pine, $r^2=0.85$, $P<0.001$) indicating that elevated CO₂ levels caused an increase in leaf c_i/c_a . The observed increase in $\Delta^{13}\text{C}$ values reflect a 13% and 17.5% increase in leaf c_i/c_a in the case of toadflax and lodgepole pine samples, respectively.

We observed a 2.4‰ decrease in $\delta^{18}\text{O}$ of leaf biomass with increasing CO₂ exposure in dalmation toadflax and a 2.5‰ increase in $\delta^{18}\text{O}$ of leaf biomass along the gradient of increasing CO₂ exposure in lodgepole pine (Fig. 3a and b). Because of close proximity, we assumed that the oxygen isotopic composition of soil water and atmospheric vapor was similar for plants growing within control or elevated CO₂ sites, and the conceptual model proposed by Grams et al. (2007) can be used to understand components of the photosynthetic response of the two species. This model infers the changes in stomatal conductance, c_i/c_a and photosynthetic capacity of plants using a dual carbon and oxygen isotope approach. This model is an expansion of the conceptual model proposed by Scheidegger et al. (2000) which estimates changes in leaf-to-air vapor pressure deficit from the $\delta^{18}\text{O}$ of leaf biomass for prediction of changes that would have likely occurred in stomatal conductance. The model of Grams et al. (2007) infers stomatal conductance changes directly from $\delta^{18}\text{O}$ of leaf cellulose bypassing the previously used relative humidity/ $\delta^{18}\text{O}$ relationship in the model of Scheidegger et al. (2000). Grams et al. (2007) did not find a significant correlation between $\delta^{18}\text{O}$ of leaf biomass and leaf cellulose in their study and therefore used the $\delta^{18}\text{O}$ of leaf cellulose as the input parameter in their model. However, a reliable correlation between $\delta^{18}\text{O}$ of leaf biomass and cellulose has been

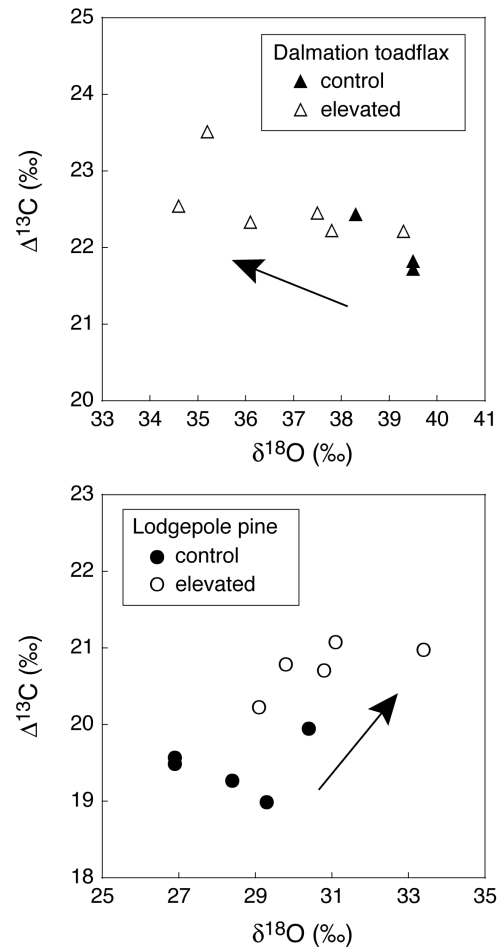


Fig. 3. Correlation between ¹³C discrimination ($\Delta^{13}\text{C}$) and the $\delta^{18}\text{O}$ of bulk leaf material in (a) dalmation toadflax and (b) lodgepole pine. The filled and open symbols represent data from the control and elevated sites respectively.

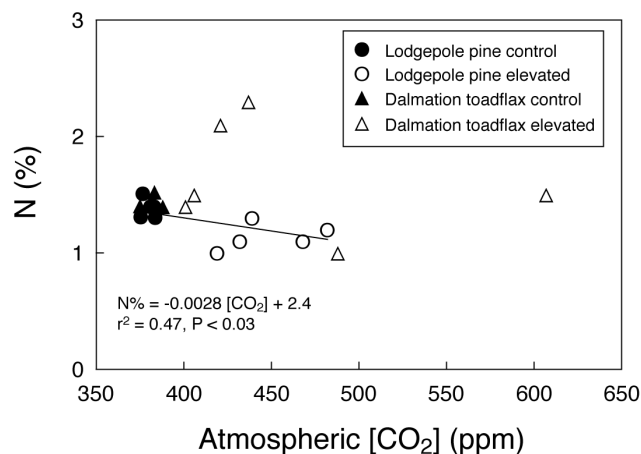
observed in several herbaceous and woody plants (Barbour and Farquhar 2000; Barbour et al., 2001). We feel justified in using the $\delta^{18}\text{O}$ of leaf biomass instead of $\delta^{18}\text{O}$ of leaf cellulose as an input parameter in the conceptual model of Grams et al. (2007).

In case of dalmation toadflax there is an increase in $\Delta^{13}\text{C}$ accompanied by a decrease in $\delta^{18}\text{O}$ of leaf biomass associated with increasing CO₂ exposure (Fig. 3a). The decrease in $\delta^{18}\text{O}$ of leaf biomass is likely a result of an increase in stomatal conductance. This is because higher stomatal conductance results in a higher rate of transpiration which cools the leaf surface and reduces the leaf intercellular vapor pressure resulting in lower $\delta^{18}\text{O}$ enrichment at evaporating sites. Further, due to increased transpiration, the convective flux of unenriched source water to evaporating sites increases, thereby reducing the enrichment of bulk leaf water (Farquhar and Lloyd 1993). This $\delta^{18}\text{O}$ enrichment signal of leaf water is transferred to the bulk leaf organic matter by isotope

exchange reactions. Using the dual isotope model of Grams et al. (2007) the increase in c_i/c_a can then be inferred to be a result of increased stomatal conductance accompanied by no response of photosynthetic capacity. Our results do not fit predictions from FACE sites which generally suggest increasing exposure to CO₂ results in an average 22% decrease in stomatal conductance across all plant species (reviewed in Ainsworth and Rogers, 2007). However, preliminary studies in a CO₂ enrichment experiment (www.phace.us) near Cheyenne, Wyoming USA (Williams, unpublished data) also confirm that dalmation toadflax shows about a 6‰ ($n=2-3$) decline in leaf biomass $\delta^{18}\text{O}$ values under elevated CO₂ conditions.

Several studies have shown that soil water savings induced by decreased stomatal conductance under elevated CO₂ conditions is primarily observed when moisture is limiting (Field et al., 1997; Wullschlegel et al., 2002). However, when soil water availability is high several plant species show higher stomatal conductance and are insensitive to elevated CO₂ conditions (Mooney et al., 1983; Knapp et al., 1996; Schomp, 2007). We hypothesize that since dalmation toadflax maintains deeper maximal rooting depth and greater root mass density in deep soil layers compared to other co-existing native species it has access to surplus moisture stored in deeper soil layers under elevated CO₂ conditions (Schomp, 2007). Due to this more favorable plant water status it shows higher stomatal conductance under elevated CO₂ conditions.

A strikingly different pattern was observed in lodgepole pine. Here an increase in $\Delta^{13}\text{C}$ was accompanied by an increase in $\delta^{18}\text{O}$ of leaf biomass with increasing CO₂ exposure (Fig. 3b). The increase in $\delta^{18}\text{O}$ of leaf biomass is likely a result of decreased stomatal conductance. The decreased stomatal conductance should result in a decrease in c_i/c_a if photosynthetic CO₂ demand is unchanged. However, we observed an increase in c_i/c_a in lodgepole pine exposed to elevated CO₂ levels likely due to decreased photosynthetic capacity. The N concentration of plant leaves is positively correlated with photosynthetic capacity (Field and Mooney, 1986), as most of the N in leaves is used for synthesis of photosynthetic enzymes, and in particular Rubisco (Shangguan et al., 2000). The N concentration of lodgepole pine decreases with an increase in atmospheric CO₂ concentration ($r^2=0.47$; $P<0.03$) (Fig. 4). This decrease in leaf N% at elevated CO₂ might be an additional indicator of decreased photosynthetic capacity in lodgepole pine at elevated CO₂ levels. No correlation was observed between leaf N and $\Delta^{13}\text{C}$ in dalmation toadflax (Fig. 4). Therefore we can infer that photosynthetic capacity of lodgepole pine decreases under higher CO₂ concentrations. Our result again contradicts the results obtained from FACE sites which frequently show that elevated CO₂ stimulates photosynthesis in C3 plants, with trees showing the largest net increase in photosynthesis with increasing CO₂ exposure (reviewed in Nowak et al., 2004; Ainsworth and Long, 2005).



such studies direct measurement of leaf gas exchange rates, stomatal conductance and the response of photosynthesis to leaf internal CO₂ concentration (A-ci curves) should also be used in conjunction to isotopes to determine direct short-term responses of these plants to elevated CO₂.

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