



Modelling the impact of riverine DON removal by marine bacterioplankton on primary production in the Arctic Ocean

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Abstract. The planktonic and biogeochemical dynamics of the Arctic shelves exhibit a strong variability in response to Arctic warming. In this study, we employ a biogeochemical model coupled to a pan-Arctic ocean–sea ice model (MIT-gcm) to elucidate the processes regulating the primary production (PP) of phytoplankton, bacterioplankton (BP), and their interactions. The model explicitly simulates and quantifies the contribution of usable dissolved organic nitrogen (DON) drained by the major circum-Arctic rivers to PP and BP in a scenario of melting sea ice (1998–2011). Model simulations suggest that, on average between 1998 and 2011, the removal of usable riverine dissolved organic nitrogen (RDON) by bacterioplankton is responsible for a $\sim 26\%$ increase in the annual BP for the whole Arctic Ocean. With respect to total PP, the model simulates an increase of $\sim 8\%$ on an annual basis and of $\sim 18\%$ in summer. Recycled ammonium is responsible for the PP increase. The recycling of RDON by bacterioplankton promotes higher BP and PP, but there is no significant temporal trend in the BP:PP ratio within the ice-free shelves over the 1998–2011 period. This suggests no significant evolution in the balance between autotrophy and heterotrophy in the last decade, with a constant annual flux of RDON into the coastal ocean, although changes in RDON supply and further reduction in sea-ice cover could potentially alter this delicate balance.

1 Introduction

In response to the polar amplification of global climate change, air temperature in the lower atmosphere is increasing twice as fast in the Arctic as in temperate regions. By the end of the century, model projections suggest an average increase in the surface air temperature of 3.7°C relative to 1981–2000 (ACIA report, 2005). In response to Arctic warming, plankton production and the biogeochemistry of the Arctic Ocean (AO) are rapidly evolving. Changes in phytoplankton communities (Li et al., 2009) as well as their phenology in spring (Kahru et al., 2011) and autumn (Ardyna et al., 2014) are being observed. Overall, the AO tends to be more productive (Bélanger et al., 2013) and is taking up more atmospheric carbon dioxide (1996–2007; Manizza et al., 2013). In the long term, model projections suggest an increase in spatially integrated primary production (PP) by the end of the twenty-first century (Vancoppenolle et al., 2013).

The AO is the basin most influenced by continental freshwater. It receives 10% of the freshwater that flows into the global ocean, but represents only 1% of the global ocean volume (Opshal et al., 1999). Circum-Arctic rivers are potentially a significant source of inorganic nutrients and organic matter for shelf seas (Le Fouest et al., 2013; Tank et al., 2012). 10% of the global riverine inputs of organic carbon are conveyed into the AO (Rachold et al., 2004). This fraction is projected to increase in the near future due to the ac-

celerated thawing of permafrost (Frey et al., 2007). This pool of organic matter enters the carbon cycle, but little is known about its fate and pathways within the plankton ecosystem in Arctic waters prior to being exported into the Atlantic Ocean.

Bacterioplankton is a major biological component involved in the degradation and mineralization of dissolved organic matter in Arctic waters (Ortega-Retuerta et al., 2012a). It can significantly affect the fate and distribution of organic matter within the entire water column (Bendsten et al., 2002) as well as the microbial food web activity through the assimilation of remineralized nitrogen. However, the contribution of Arctic bacterioplankton to plankton production in the context of Arctic warming remains unknown. Despite the fact that the AO basin now acts as a sink for atmospheric carbon dioxide (1996–2007; Manizza et al., 2013), the balance between autotrophy and heterotrophy may change in the future based on observations of enhanced stratification of the water column (Li et al., 2009), increased sea temperature (Timmermans et al., 2014; Steele et al., 2008), which acts as a key driver of Arctic bacterioplankton metabolism (Piontek et al., 2014; Bendsten et al., 2002), and changes in the riverine inputs of nutrients due to an increase in freshwater discharge (Shiklomanov and Lammers, 2011). In near-shore AO waters, riverine inputs already sustain part of the bacterial activity (e.g. Vallières et al., 2008).

Using a relatively simple biogeochemical modelling approach, Tank et al. (2012) shed light on the potential impact of riverine nutrient inputs on the PP of the AO. In the present study, we propose building on the static view provided by the work of Tank et al. (2012) by explicitly modelling the effect of the interactions between riverine dissolved organic nitrogen (RDON) and bacterioplankton. The objective is to use a pan-Arctic ocean–sea ice coupled model to quantify the contribution of usable RDON processed by marine bacterioplankton to the production of both bacterioplankton and phytoplankton in a scenario of melting sea ice over the period 1998–2011.

2 Material and methods

2.1 The physical model

We used the MIT general circulation model (MITgcm) (Marshall et al., 1997) coupled with a sea-ice model. The model is configured on a “cubed-sphere” grid encompassing the Arctic domain with open boundaries at $\approx 55^\circ$ N in the Atlantic and Pacific sectors. Prescribed boundary conditions for potential temperature, salinity, flow, and sea-surface elevation are provided from previous integrations of a global configuration of the same model (Menemenlis et al., 2005). The grid has a variable horizontal resolution with an average mesh of ~ 18 km. The mesh resolves major Arctic straits, including many of the channels of the Canadian Archipelago. The sea-ice and fluid dynamics equations are solved on the same

horizontal mesh. The 50-level vertical grid is height based, varying from 10 m thick near the surface to ~ 450 m at a depth of ~ 6 km. Bathymetry is derived from the US National Geophysical Data Center (NGDC) two-minute global relief data set (ETOPO2), which uses the International Bathymetric Chart of the Arctic Ocean (IBCAO) product for Arctic bathymetry (Jakobsson et al., 2008). The ETOPO2 data are smoothed to the model’s horizontal mesh and mapped to the ocean’s vertical levels using a “lopped cell” strategy (Adcroft et al., 1997), which permits an accurate representation of the ocean bottom boundary.

The ocean model’s hydrography is initialized with observations taken from the Polar Science Center Hydrographic Climatology (PHC) 3.0 database (Steele et al., 2001). Initial sea-ice distributions are taken from the pan-Arctic Ice-Ocean Modeling and Assimilation System data sets (Zhang and Rothrock, 2003). Atmospheric forcings (10 m surface winds, 2 m air temperature and humidity, and downward longwave and shortwave radiation) are taken from the 6-hourly data sets of the Japanese 25-year ReAnalysis (JRA-25; Onogi et al., 2007). Monthly mean estuarine fluxes of freshwater are based on the Arctic Runoff database (Lammers et al., 2001; Shiklomanov et al., 2000). The sea-ice component of the coupled model follows the viscous-plastic rheology formulation of Hibler (1979) with momentum equations solved implicitly on a C-grid (Arakawa and Lamb, 1977) using a procedure based on Zhang and Hibler (1997). Fluxes of momentum into ice due to the overlying atmospheric winds and momentum fluxes between sea ice and the ocean are calculated by solving for the momentum balance at each surface grid column (Hibler and Bryan, 1987). This model configuration was previously used to study the Arctic freshwater budget (Condrón et al., 2009). Modelling studies of Condrón et al. (2009) compared to observations by Serreze et al. (2006) concluded that this model configuration is able to realistically represent the freshwater budget of the AO, including the import and export of freshwater from the Bering and Fram straits and from the Canadian Archipelago.

2.2 The riverine DON (RDON) discharge

To realistically represent the RDON flux in the AO in our biogeochemical model, we follow the approach adopted by Manizza et al. (2009), which is based on seasonally explicit regression relationships. These relationships use co-variations between water yield and dissolved organic carbon (DOC) concentrations in circum-Arctic rivers to define riverine DOC (RDOC) monthly averaged fluxes for 10 regions in the pan-Arctic domain. These regions are the Barents Sea, Kara Sea, Laptev Sea, East Siberian Sea, Chukchi Sea, Bering Strait, Beaufort Sea, Canadian Archipelago, Hudson Bay, and Hudson Strait using published watershed areas and seasonal water runoff (Lammers et al., 2001). The approach uses empirical relationships quantifying the co-variation between discharge and RDOC to scale the Lam-

mers et al. (2001) discharge estimates into estimates of RDOC export. Estimates of RDOC export for December–March, April–July, and August–November were divided into monthly bins according to measured distributions of RDOC export for those months in Arctic rivers. For each season, [RDOC]–discharge relationships were developed. North American and Eurasian rivers were considered separately. Data from the Yukon, Mackenzie, and Kuparuk rivers were used to define a runoff–[RDOC] relationship for drainage areas in North America, and data from the Ob', Yenisey, and Lena rivers were used to define a runoff–[RDOC] relationship for drainage areas in Eurasia. RDOC for the Yenisey, Ob', Lena, and Mackenzie were collected as part of the Pan-Arctic River Transport of Nutrients, Organic Matter, and Suspended Sediments (PARTNERS) project (McClelland et al., 2008). RDOC concentrations for the Kuparuk River were collected as part of the NSF Study of the Northern Alaska Coastal System (SNACS, <http://www.arcus.org/arcss/snacs/index.php>). In all cases, discharge data were acquired from ArcticRIMS (<http://rims.unh.edu/>). Recent sampling efforts on these rivers have provided exceptional seasonal coverage (McClelland et al., 2008) and the total annual discharge of RDOC in the model is 37.7 TgC yr^{-1} , which is consistent with the estimate of Raymond et al. (2007). To initialize the model, we used the three-dimensional RDOC field obtained from the 3-decade integration of the model by Manizza et al. (2009). After that time, RDOC distributions are relatively steady, because the flushing time for tracers through the surface waters of the basin is of the order of a decade. RDOC was converted into nitrogen currency (RDON) using a molar C : N ratio of 40 : 1 (Tank et al., 2012; Köhler et al., 2003). We assume that 15 % of the RDON entering the model at river grid cells is usable by bacterioplankton (e.g. Wickland et al., 2012).

2.3 The biogeochemical model

We couple to the MITgcm physical model a biogeochemical model that explicitly represents the plankton ecosystem dynamics. The biogeochemical model is improved from previous applications in sub-Arctic (Le Fouest et al., 2005, 2006) and Arctic waters (Le Fouest et al., 2011, 2013b). In the model, nitrogen is the currency and it includes 10 compartments (i.e. nine in the pelagic domain + RDON that couples the marine and terrestrial cycling of nitrogen), chosen according to the ecosystem structure observed in the AO. Phytoplankton is size-fractionated into large ($> 5 \mu\text{m}$) and small ($< 5 \mu\text{m}$) phytoplankton (LP and SP, respectively). These two compartments encompass the major phytoplankton groups relevant for plankton dynamics and biogeochemistry in the Arctic waters (e.g. Li et al., 2009; Coupel et al., 2012). The two zooplankton compartments represent large (LZ, mainly copepods) and small (SZ, protozooplankton) organisms. Dissolved inorganic nutrients are nitrate (NO_3) and ammonium (NH_4). Detrital (i.e. produced by the bio-

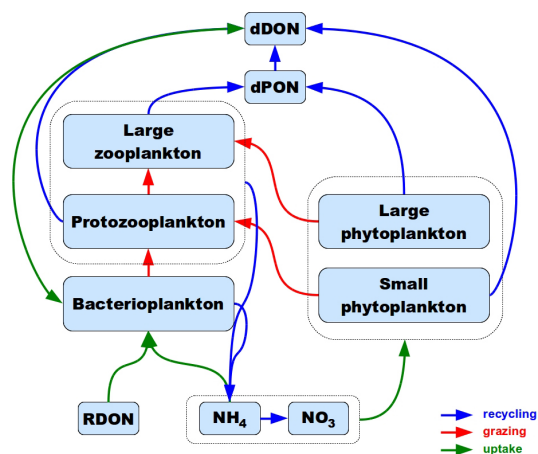


Figure 1. Conceptual diagram of the biogeochemical model. The 10 state variables are nitrate (NO_3), ammonium (NH_4) and small ($< 5 \mu\text{m}$) phytoplankton, large zooplankton, protozooplankton, bacterioplankton, detrital particulate and dissolved organic nitrogen (dPON and dDON, respectively), and usable riverine dissolved organic nitrogen (RDON). Green, red and blue arrows represent nutrient uptake, grazing, and nitrogen recycling, respectively.

geochemical model components) particulate and dissolved organic nitrogen (dPON and dDON, respectively) close the nitrogen cycle. Bacterioplankton (BACT) are explicitly represented following the model of Fasham et al. (1990). They grow on NH_4 , dDON, and on the usable fraction of RDON (see the Appendix for details). LP and SP growth depends on light, NO_3 and NH_4 availability according to Liebig's law of the minimum. LZ graze on LP and SZ, whereas SZ graze on SP and BACT. Fecal pellets and LP basal mortality fuel the dPON pool. The dDON pool is made of unassimilated nitrogen resulting from SZ grazing, SP, SZ and BACT basal mortality and dPON fragmentation. BACT release, LZ excretion and unassimilated nitrogen resulting from SZ grazing are the sources of NH_4 in the model. NH_4 is converted into NO_3 through the nitrification process. For phytoplankton, nitrogen is converted into carbon using the Redfield carbon to nitrogen (C : N) molar ratio (106 : 16; Redfield et al., 1963) and into Chl using variable C : Chl mass ratios computed according to a modified version of the phytoplankton photoacclimation model of Cloern et al. (1995). The plankton biogeochemical model (Fig. 1) is fully detailed in the Appendix. Differential equations are given in Table 1, whereas biological parameters are given in Table 2.

Nitrate data used for the model initialization are from the World Ocean Atlas 2005 (National Oceanographic Data Centre, 2006). LP and SP are assigned a constant field over the model grid (0.2 and $0.002 \text{ mmol N m}^{-3}$ in the top eight layers and below, respectively; e.g. Sherr et al., 2003; Ducklow, 1999, Taniguchi, 1999). The same conditions are imposed for BACT (e.g. Sherr et al., 2003; Ducklow, 1999).

Table 1. Differential equations for the 10-component biogeochemical model: nitrate (NO_3), ammonium (NH_4), large and small phytoplankton (LP and SP, respectively), large and small zooplankton (LZ and SZ, respectively), bacterioplankton (BACT), detrital particulate and dissolved organic nitrogen (dPON and dDON, respectively), and usable riverine dissolved organic nitrogen (RDON).

$$\begin{aligned} \frac{\partial \text{NO}_3}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{NO}_3 - \mathbf{K} \cdot \nabla \text{NO}_3) + \text{nitrif} - \lim_{\text{NO}_3}^{\text{LP}} \mu_{\text{LP}} \text{LP} - \lim_{\text{NO}_3}^{\text{SP}} \mu_{\text{SP}} \text{SP} \\ \frac{\partial \text{NH}_4}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{NH}_4 - \mathbf{K} \cdot \nabla \text{NH}_4) - \lim_{\text{NH}_4}^{\text{LP}} \mu_{\text{LP}} \text{LP} - \lim_{\text{NH}_4}^{\text{SP}} \mu_{\text{SP}} \text{SP} - \text{nitrif} - \text{Ubact}_{\text{NH}_4} \text{BACT} (1 - \text{ge}_{\text{BACT}}) + \text{eg}_{\text{SZ}} (1 - \text{ge}_{\text{SZ}}) G_{\text{SZ}} \text{SZ} + \text{ex}_{\text{LZ}} \text{LZ} \\ \frac{\partial \text{LP}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{LP} - \mathbf{K} \cdot \nabla \text{LP}) + \mu_{\text{LP}} \text{LP} - G_{\text{LZ}} \text{pf}_{\text{LP}} \text{LZ} - m_{\text{LP}} \text{LP} + \frac{\partial}{\partial z} (\text{sed}_{\text{LP}} \text{LP}) \\ \frac{\partial \text{SP}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{SP} - \mathbf{K} \cdot \nabla \text{SP}) + \mu_{\text{SP}} \text{SP} - G_{\text{SZ}} \text{pf}_{\text{SP}} \text{SZ} - m_{\text{SP}} \text{SP} \\ \frac{\partial \text{LZ}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{LZ} - \mathbf{K} \cdot \nabla \text{LZ}) + \text{assim}_{\text{LZ}} G_{\text{LZ}} \text{LZ} - m_{\text{LZ}} \text{LZ}^2 - \text{ex}_{\text{LZ}} \text{LZ} \\ \frac{\partial \text{SZ}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{SZ} - \mathbf{K} \cdot \nabla \text{SZ}) + \text{assim}_{\text{SZ}} G_{\text{SZ}} \text{SZ} - m_{\text{SZ}} \text{SZ}^2 - G_{\text{LZ}} (1 - \text{pf}_{\text{LP}}) \text{LZ} \\ \frac{\partial \text{BACT}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{BACT} - \mathbf{K} \cdot \nabla \text{BACT}) + \text{Ubact}_{\text{NH}_4} \text{BACT} \text{ge} + \text{Ubact}_{\text{dDON}} \text{BACT} \text{ge}_{\text{BACT}} - m_{\text{BACT}} \text{BACT} - G_{\text{SZ}} (1 - \text{pf}_{\text{SP}}) \text{SZ} \\ \frac{\partial \text{dPON}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{dPON} - \mathbf{K} \cdot \nabla \text{dPON}) + (1 - \text{assim}_{\text{LZ}}) G_{\text{LZ}} \text{LZ} + m_{\text{LZ}} \text{LZ}^2 + m_{\text{LP}} \text{LP} + \frac{\partial}{\partial z} (\text{sed}_{\text{dpon}} \text{dPON}) - f_{\text{g}} \text{dPON} \\ \frac{\partial \text{dDON}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{dDON} - \mathbf{K} \cdot \nabla \text{dDON}) + f_{\text{g}} \text{dPON} + m_{\text{SZ}} \text{SZ}^2 + m_{\text{SP}} \text{SP} + m_{\text{BACT}} \text{BACT} + (1 - \text{eg}_{\text{SZ}}) (1 - \text{ge}_{\text{SZ}}) G_{\text{SZ}} \text{SZ} - \text{Ubact}_{\text{DON}} \text{BACT} \text{pf}_{\text{DON}} (1 - \text{ge}_{\text{BACT}}) \\ \frac{\partial \text{RDON}}{\partial t} &= -\nabla \cdot (\mathbf{u}\text{RDON} - \mathbf{K} \cdot \nabla \text{RDON}) - \text{Ubact}_{\text{DON}} \text{BACT} (1 - \text{pf}_{\text{DON}}) (1/\text{ratio}_{\text{CN}}) (1 - \text{ge}_{\text{BACT}}) \end{aligned}$$

LZ and SZ are assigned a constant field over the model grid (0.1 and 0.001 mmol N m⁻³ in the top eight layers and below, respectively; e.g. Sherr et al., 2003, Taniguchi, 1999). The same conditions are imposed a priori for dPON. A value of 1 mmol N m⁻³ of NH₄ (e.g. Kristiansen et al., 1994) and dDON (e.g. Charria et al., 2008) is imposed at each grid cell. Boundary conditions at the North Atlantic and North Pacific sectors are data from the World Ocean Atlas 2005 (NODC, 2006) for NO₃, and null for the remaining nine biogeochemical tracers. Apart from RDON, there are no riverine inputs for the remaining nine biogeochemical tracers.

2.4 Coupled model integrations

The model is spun up by repeating the January 1980–December 1989 decade twice. It is thereafter initialized with the physical and biogeochemical fields obtained from 31 December 1989 to run the 1990–2011 time period. Two simulations are then carried out: without usable RDON removal by bacterioplankton (our control run, hereafter CTRL run) and with usable RDON removal by bacterioplankton (hereafter RIV run). The difference between the two simulations provides information on the potential impact of the interactions between bacterioplankton and usable RDON on bacterioplankton production (BP), nutrient regeneration, and ultimately primary production (PP) in the Arctic basin.

3 Results

3.1 Primary production

Shelf seas influenced least by riverine inputs of RDON show comparable simulated annual rates of total PP in the CTRL and RIV runs (Fig. 2). In the Barents Sea, simulated PP averaged over 1998–2011 reaches up to ~80 gC m⁻² yr⁻¹, in line with previous remote sensing estimates (up to 70–80 gC m⁻² yr⁻¹ on average over 1998–2010; Bélanger et al., 2013). In the central Chukchi Sea, simulated PP lies between 50 and 80 gC m⁻² yr⁻¹, in agreement with the observed range (15–80 gC m⁻² yr⁻¹ on average over 1998–2007; Bélanger et al., 2013).

The largest differences in total PP between the two runs are found in the river-influenced Eurasian seas (East Siberian Sea, Laptev Sea, and Kara Sea; Fig. 2). In the CTRL run, maximum simulated PP rates reach ~30 gC m⁻² yr⁻¹, which is more than 3-fold lower than satellite-derived and in situ estimates that can exceed 100 gC m⁻² yr⁻¹ (Bélanger et al., 2013; Codispoti et al., 2013; Sakshaug, 2004). In contrast, PP rates simulated in the RIV run (80–90 gC m⁻² yr⁻¹) show a better agreement with observations.

The increase in the 1998–2011 averaged annual PP in the RIV run relative to the CTRL run is due to the increase in NH₄-supported PP (Fig. 3d, e and f). In contrast, overall, new PP remains unaffected by the bacterial use of RDON (Fig. 3a, b and c). In the Kara Sea, Laptev

Table 2. Biogeochemical model parameters.

Symbol	Description	Value	Units
Nutrients			
nitrif_{\max}	Maximum NH_4 nitrification rate	0.05	d^{-1}
K_{nitrif}^N	Half-saturation constant for NH_4 nitrification	0.07	mmol N m^{-3}
Phytoplankton			
k_w	Light attenuation coefficient due to water	0.04	m^{-1}
k_{nonchl}	Light attenuation coefficient due to nonchlorophyllous matter	0.05	m^{-1}
$K_{\text{NO}_3}^{\text{LP}}$	Half-saturation constant for NO_3 use by LP	1	mmol N m^{-3}
$K_{\text{NO}_3}^{\text{SP}}$	Half-saturation constant for NO_3 use by SP	1	mmol N m^{-3}
$K_{\text{NH}_4}^{\text{LP}}$	Half-saturation constant for NH_4 use by LP	0.5	mmol N m^{-3}
$K_{\text{NH}_4}^{\text{SP}}$	Half-saturation constant for NH_4 use by SP	0.1	mmol N m^{-3}
K_E^{LP}	Photoacclimation parameter	8	$\text{Ein m}^{-2} \text{d}^{-1}$
K_E^{SP}	Photoacclimation parameter	4	$\text{Ein m}^{-2} \text{d}^{-1}$
$\left(\frac{\text{Chl}}{\text{C}}\right)_{\text{LP}}^{\max}$	Maximum Chl to C ratio for LP	0.0125	g g^{-1}
$\left(\frac{\text{Chl}}{\text{C}}\right)_{\text{SP}}^{\max}$	Maximum Chl to C ratio for SP	0.07	g g^{-1}
μ_{\max}^{LP}	Maximum growth rate for LP	1.4	d^{-1}
μ_{\max}^{SP}	Maximum growth rate for SP	1.4	d^{-1}
α_{SP}	Initial slope of the photosynthesis–irradiance curve	5.5	$\text{mg C (mg Chl)}^{-1} (\text{Ein m}^{-2} \text{d}^{-1})^{-1}$
α_{LP}	Initial slope of the photosynthesis–irradiance curve	7.5	$\text{mg C (mg Chl)}^{-1} (\text{Ein m}^{-2} \text{d}^{-1})^{-1}$
sed_{lp}	LP sinking rate	2	m^{-1}
m_{LP}	LP basal mortality	0.05	d^{-1}
m_{SP}	SP basal mortality	0.05	d^{-1}
Zooplankton			
G_{LZ}^{\max}	Maximum grazing rate for LZ	0.3	d^{-1}
λ	Ivlev constant for LZ	0.5	$(\text{mmol N m}^{-3})^{-1}$
G_{SZ}^{\max}	Maximum grazing rate for SZ	1	d^{-1}
K_G	Half-saturation constant for SZ grazing	0.8	mmol N m^{-3}
assim_{LZ}	LZ assimilation efficiency	70	%
ge_{SZ}	SZ growth efficiency	30	%
eg_{SZ}	dDON egestion by SZ	40	%
ex_{LZ}	NH_4 excretion by LZ	0.05	d^{-1}
m_{SZ}	SZ mortality	0.05	$(\text{mmol N m}^{-3})^{-1}$
m_{LZ}	LZ mortality	0.2	$(\text{mmol N m}^{-3})^{-1}$
Bacterioplankton			
U_{bact}_{\max}	Maximum growth rate	1	d^{-1}
$K_{\text{NH}_4}^{\text{BACT}}$	Half-saturation constant for NH_4 uptake	0.1	mmol N m^{-3}
$K_{\text{DON}}^{\text{BACT}}$	Half-saturation constant for DON uptake	0.1	mmol N m^{-3}
ge_{BACT}	Growth efficiency	20	%
m_{BACT}	Basal mortality	0.05	d^{-1}
Detritus			
sed_{dpon}	dPON sinking rate	100	$\text{m d}^{-1} (\text{mmol N m}^{-3})^{-1}$
f_g	dPON fragmentation	0.05	d^{-1}

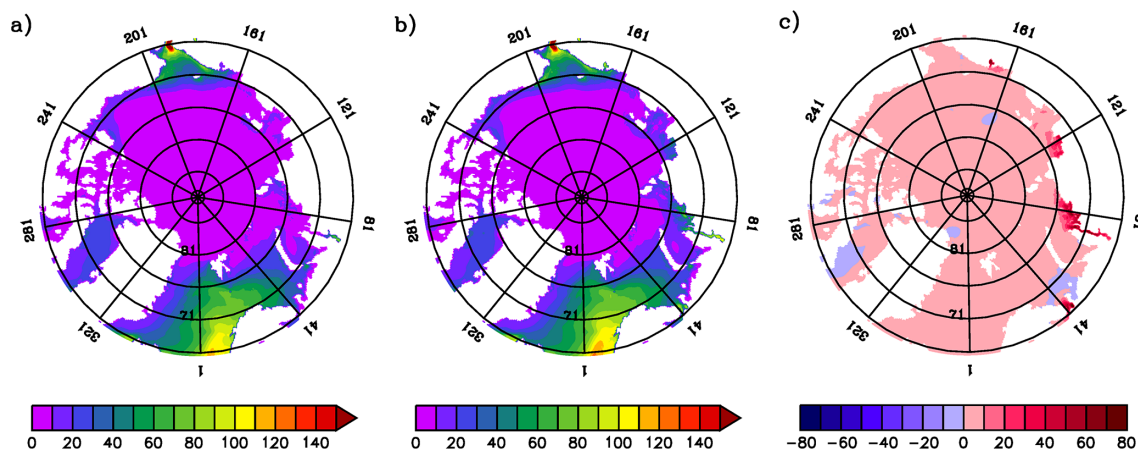


Figure 2. Mean annual ocean primary production (gC m^{-2}) over 1998–2011 (a) without RDON removal by bacterioplankton (CTRL run) and (b) with RDON removal by bacterioplankton (RIV run), and (c) the absolute difference (gC m^{-2} ; RIV run – CTRL run).

Sea, East Siberian Sea, and Beaufort Sea, simulated new PP is mostly $< 20 \text{ gC m}^{-2} \text{ yr}^{-1}$, in agreement with previously estimated rates ($< 17 \text{ gC m}^{-2} \text{ yr}^{-1}$; Sakshaug, 2004). New PP rates simulated by the model in the more productive areas are also in line with Sakshaug's estimated rates. In the Chukchi Sea, new PP generally lies in the $10\text{--}30 \text{ gC m}^{-2} \text{ yr}^{-1}$ range and reaches $> 100 \text{ gC m}^{-2} \text{ yr}^{-1}$ at the sea opening ($5\text{--}160 \text{ gC m}^{-2} \text{ yr}^{-1}$; Sakshaug, 2004). Simulated new PP is up to $\sim 70 \text{ gC m}^{-2} \text{ yr}^{-1}$ in the Barents Sea, close to the value given by Sakshaug (up to $100 \text{ gC m}^{-2} \text{ yr}^{-1}$; 2004). In the Greenland and Labrador seas, the simulated new PP rates are ~ 50 and $\sim 30 \text{ gC m}^{-2} \text{ yr}^{-1}$, respectively ($40\text{--}45 \text{ gC m}^{-2} \text{ yr}^{-1}$; Sakshaug, 2004).

Direct estimates of NH_4 -supported PP based on measurements are rare in Arctic coastal waters. Nevertheless, they can be crudely derived by subtracting new PP from total PP estimated by Sakshaug (2004). In the Eurasian and North American shelves, NH_4 -supported PP in the CTRL run is $< 10 \text{ gC m}^{-2} \text{ yr}^{-1}$ (Fig. 3d) overall. This is low relative to the rates derived from Sakshaug's data, which would range from ~ 25 to $\sim 40 \text{ gC m}^{-2} \text{ yr}^{-1}$. By contrast, in the RIV run, rates simulated in offshore shelf waters are $\sim 10\text{--}30 \text{ gC m}^{-2} \text{ yr}^{-1}$. However, closer to the coast, local rates reach $40\text{--}50$ (Laptev Sea) and $70\text{--}80 \text{ gC m}^{-2} \text{ yr}^{-1}$ (Kara Sea; Fig. 3e).

Averaged over 1998–2011, the total PP simulated by the model and integrated over the whole AO is $662 \pm 91 \text{ TgC yr}^{-1}$ in the CTRL run and $717 \pm 95 \text{ TgC yr}^{-1}$ in the RIV run. These values are within the range of previously reported rates based on remote sensing or in situ data ($385\text{--}1008 \text{ TgC yr}^{-1}$, Bélanger et al., 2013; Codispoti et al., 2013; Hill et al., 2013; Arrigo and van Dijken, 2011). Between the two model runs, the annual total PP increased by $\sim 8\%$, on average, between 1998 and 2011. In September–October, when the simulated sea-ice concentration reaches its seasonal minimum, the annual total PP increase is more than twice this value ($\sim 18\%$, on average).

3.2 Bacterioplankton activity

The PP increase is tightly linked to a higher bacterioplankton activity that promotes RDON recycling into nutrients usable by both phytoplankton and bacterioplankton. The bacterioplankton biomass (BB), integrated between the sea surface and 50 m and averaged over April–June (spring) and July–September (summer), is shown in Fig. 4. As for PP, the Barents and Chukchi seas show comparable levels of BB in CTRL and RIV runs. In the Chukchi Sea, the BB simulated in spring ($< 100\text{--}250 \text{ mgC m}^{-2}$; Fig. 4a and b) overlaps with the measured range ($222\text{--}358 \text{ mgC m}^{-2}$; Kirchman et al., 2009). It is similar in summer, when simulated ($\sim 100 \text{ mgC m}^{-2}$ to $> 800 \text{ mgC m}^{-2}$; Fig. 4d and e) and measured BB levels ($250\text{--}507 \text{ mgC m}^{-2}$; Kirchman et al., 2009; Steward et al., 1996) are higher than in spring. In the Barents Sea, the simulated BB increases from $< 100 \text{ mgC m}^{-2}$ in spring to $< 250 \text{ mgC m}^{-2}$ in summer, falling within the measured range (from $\sim 80 \text{ mgC m}^{-2}$ in spring to $\sim 400 \text{ mgC m}^{-2}$ in summer, on average; Sturluson et al., 2008). In the highly river-influenced shelf seas, the two runs show notable differences in their simulated BB (Fig. 4c and f). In the central part of the Kara Sea, influenced by the Ob' and Yenisey river plumes, BB measured in late summer along a south–north transect from the Yama Peninsula to the Novaya Zemlya island is reported to range from ~ 0.1 to 7 mgC m^{-3} (Sazhin et al., 2010). For the same time period and along a comparable transect, simulated values of BB are $< 2 \text{ mgC m}^{-2}$ in the CTRL run. However, in the RIV run, BB increases up to $\sim 6\text{--}7 \text{ mgC m}^{-3}$ to match the values measured by Sazhin et al. (2010).

The depth-integrated (0–50 m) bacterioplankton production (BP) simulated in both the CTRL and RIV runs in summer in the Chukchi Sea ($< 280 \text{ mgC m}^{-2} \text{ d}^{-1}$) is consistent with measurements reported for the same season ($5\text{--}301 \text{ mgC m}^{-2} \text{ d}^{-1}$; Kirchman et al., 2009; Rich et al., 1997;

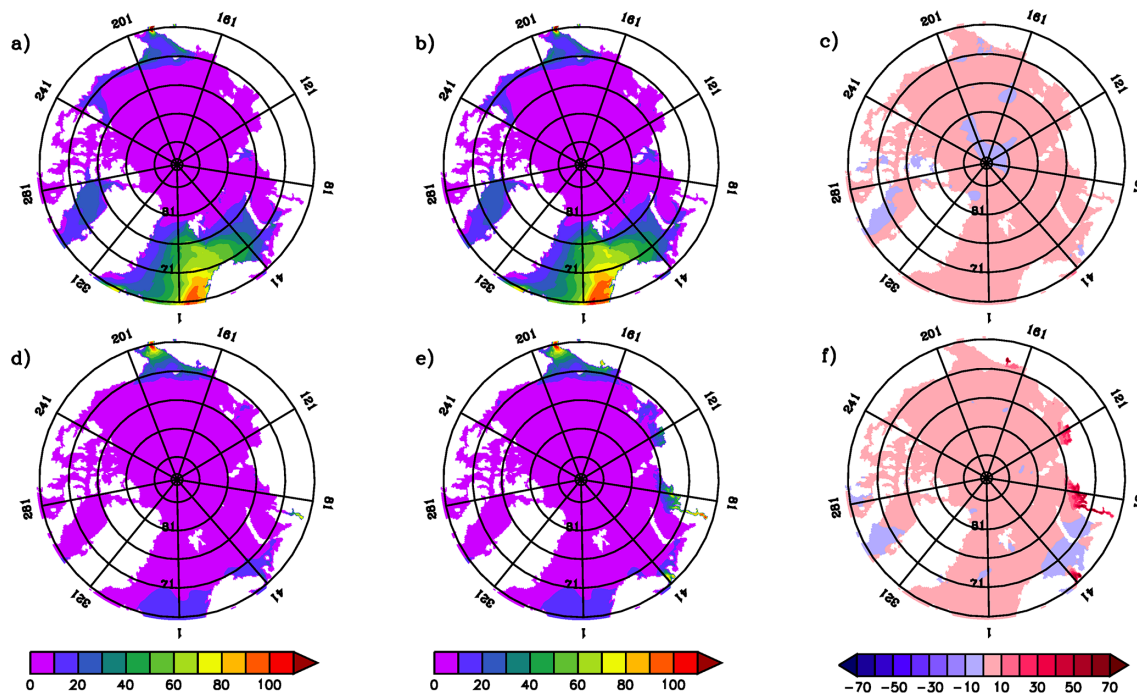


Figure 3. Mean annual new primary production (gC m^{-2} ; upper panels) and NH_4 -supported primary production (gC m^{-2} ; lower panels) over 1998–2011 simulated in the CTRL run (left panels a and d) and the RIV run (middle panels b and e). Right panels (c and f) provide the absolute difference (gC m^{-2} ; RIV run – CTRL run).

Steward et al., 1996). In the Beaufort Sea, influenced by the Mackenzie River plume, simulated BP is lower than $\sim 6 \text{ mgC m}^{-2} \text{ d}^{-1}$ in the CTRL run, which is far below measurements made within the area ($25\text{--}68 \text{ mgC m}^{-2} \text{ d}^{-1}$; Ortega-Retuerta et al., 2012a; Vallières et al., 2008). By contrast, in the RIV run, simulated BP ($< 30 \text{ mgC m}^{-2} \text{ d}^{-1}$) approaches the lower range of observations. Similarly, BP simulated in the CTRL run for the Kara Sea ($< 30 \text{ mgC m}^{-2} \text{ d}^{-1}$) does not exceed the first mid-range of measurements given by Meon and Amon (2004; $12\text{--}79 \text{ mgC m}^{-2} \text{ d}^{-1}$). In the RIV run, the simulated BP ($\sim 4\text{--}90 \text{ mgC m}^{-2} \text{ d}^{-1}$) overlaps the measured range ($12\text{--}79 \text{ mgC m}^{-2} \text{ d}^{-1}$; Meon and Amon, 2004) to reach up to $120 \text{ mgC m}^{-2} \text{ d}^{-1}$ locally. This result is consistent with enrichment experiments conducted with surface oceanic water sampled in the Beaufort Sea that showed a 43 % increase in BP when Mackenzie River water was included in samples (see Ortega-Retuerta et al., 2012a).

Averaged over 1998–2011, the total BP simulated by the model and integrated over the whole AO is, on average, 26 % higher in the RIV run ($68 \pm 9 \text{ TgC yr}^{-1}$) than in the CTRL run ($54 \pm 8 \text{ TgC yr}^{-1}$). Bacterioplankton recycle RDON into nutrients that can be used by both phytoplankton and bacterioplankton, hence promoting their growth. In addition, bacterioplankton and small phytoplankton are grazed by microzooplankton that, in turn, are grazed by mesozooplankton. More organic matter is channelled towards the upper trophic levels, a flow that also contributes to fuelling the dDON and

NH_4 pools through recycling. By enabling the removal of RDON by bacterioplankton in the biogeochemical model, the biomass of microzooplankton and mesozooplankton, averaged over 1998–2011, increased by ~ 16.1 and 43.6 %, respectively.

3.3 The bacterioplankton production versus primary production ratio (BP : PP)

The BP : PP ratio is computed over the AO shelf, delimited here by the 200 m isobaths, for ice-free waters (i.e. with less than 30 % ice cover). On average for the 1998–2011 period, the simulated BP : PP ratio is 0.19 ± 0.02 in the CTRL run and 0.21 ± 0.01 in the RIV run. These values lie within the range observed in open (0.02 ; Kirchman et al., 2009) and coastal ($0.37\text{--}0.43$; Ortega-Retuerta et al., 2012a; Garneau et al., 2008) waters. When looking at the temporal evolution of BP : PP in the RIV run (Fig. 5), the model simulates a significant increase in PP ($r = 0.57$, $p < 0.05$) and BP ($r = 0.63$, $p < 0.05$) between 1998 and 2011, with a production maximum simulated in 2007, the year showing the higher sea-ice minimum. However, there is no evidence of a significant temporal trend of BP : PP ($r = -0.09$, $p > 0.05$) over 1998–2011. This result suggests that, with a constant annual flux of RDON into the coastal AO, the significant increase in simulated BP in the model is not high enough to promote a higher contribution of heterotrophy with respect to autotrophy within the upper water column.

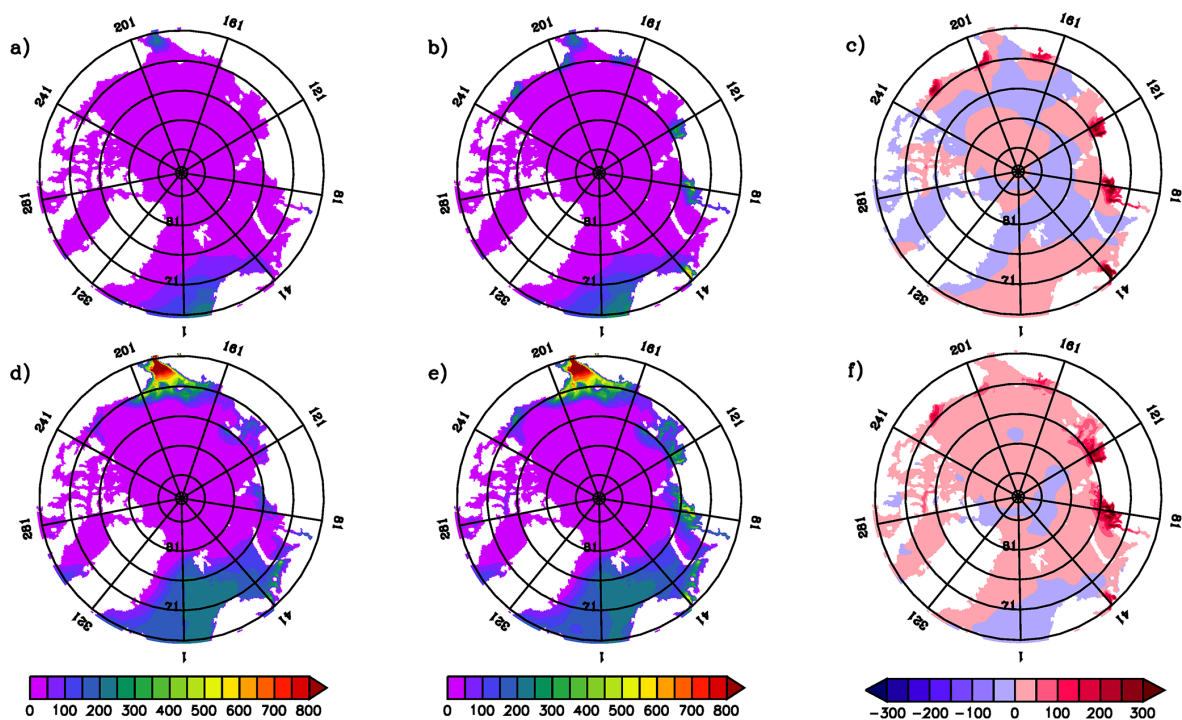


Figure 4. Seasonal climatology of the 0–50 m integrated bacterial biomass (mmolN m^{-2}) for spring (upper panels) and summer (lower panels) over the 1998–2011 period simulated in the CTRL run (left panels **a** and **d**) and in the RIV run (middle panels **b** and **e**). Right panels (**c** and **f**) provide the absolute difference (gC m^{-2} ; RIV run – CTRL run).

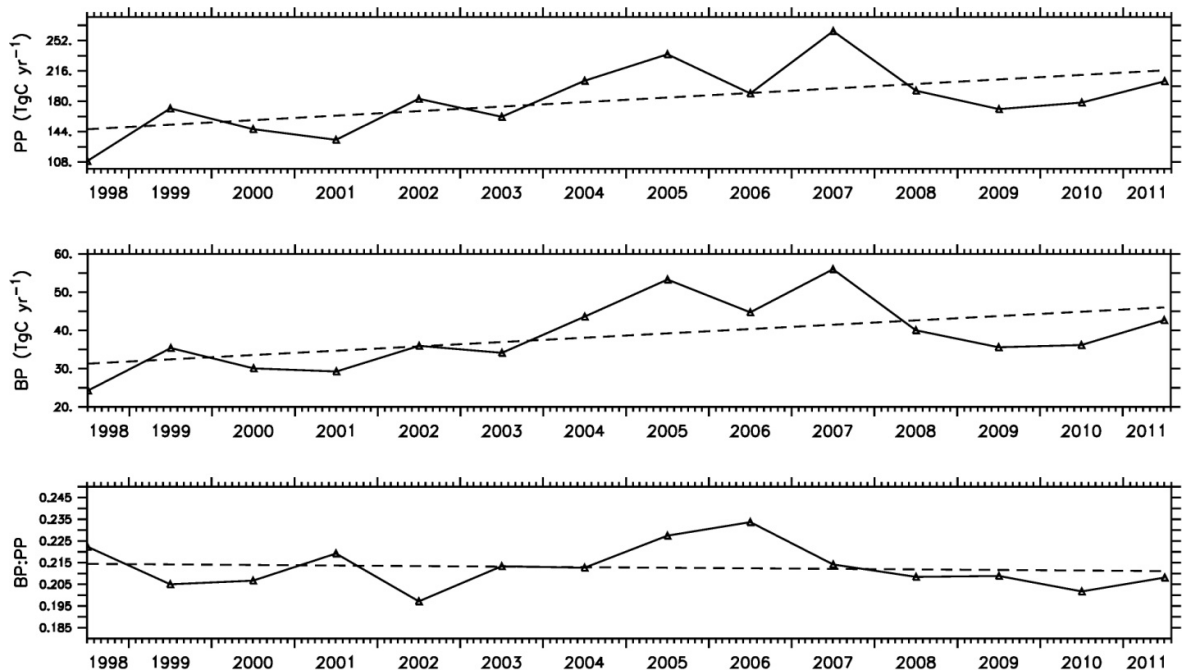


Figure 5. Time course of primary production (PP, TgC yr^{-1}) (top panel), bacterioplankton production (BP, TgC yr^{-1}) (middle panel), and the BP : PP ratio in the ice-free shelves (see text for details) of the Arctic Ocean domain ($>66.5^\circ \text{N}$) simulated in the RIV run. The dashed straight lines represent the linear trend computed over the 1998–2011 period.

4 Discussion

The coupled model suggests that NH_4 produced from the remineralization of RDON by the microbial food web contributed $\sim 8\%$ to annual pan-Arctic PP over the 1998–2011 period. This is about twice the value given in the study by Tank et al. (2012) that, in addition to RDON, accounted for the contribution of riverine inorganic nutrients as well as of the photochemical transformation of RDON into NH_4 . In our coupled model, the uptake of RDON by marine bacterioplankton and its subsequent recycling into reduced nitrogen is the sink term that shapes, with ocean transport, the spatial and temporal distribution of RDON. The photoammonification process is not parameterized but, if so, it would fuel the stock of NH_4 available for phytoplankton and bacterioplankton use, particularly in summer (e.g. Le Fouest et al., 2013; Xie et al., 2012). The RDON contribution to plankton production simulated by the coupled model can thus be considered as a minimum estimate.

From the total input of RDON, only a fraction is directly usable by the plankton (e.g. Wickland et al., 2012). The fraction that enters the coupled model by the 10 river source points is set to 15% of the total RDON input according to a study by Wickland et al. (2012), which suggests that about 15% of the total RDON pool can be degraded within less than 1 month. This value was chosen based on annual averages calculated from measurements or from model outputs for the Mackenzie River, Yukon River, Kolyma River, Lena River, Yenisey River, and Ob' River (e.g. Wickland et al., 2012). Note, however, that the average values given in Wickland et al. (2012) vary between seasons and rivers. They are lowest in the Lena River (8%) and highest in the Ob' River (19%). Maximum values as high as 24% of usable RDON are reported for the Ob' River. Sensitivity analyses with different parameterizations of the usable RDON fraction set amongst rivers and seasons would hence be informative on the amplitude of the PP and BP response to spatial and temporal variations of the usable RDON flux. To be robust, they should be combined with sensitivity analyses of the freshwater discharge to better constrain the RDON flux. In the Mackenzie River, strong inter-annual variations in terms of peak of discharge and maximum spring flow were observed in the last four decades (Yang et al., 2015). Nevertheless, the use a constant fraction of usable RDON as preformed in the present study provides a first-order estimation of its contribution to BP and PP that is consistent with the current state of knowledge about the RDON inputs. In addition to the usable RDON flux into coastal oceans, autochthonous sources of DONI (usable RDON + dDON) are important in fuelling BP. Despite improved BP estimates simulated in the RIV run, the rates remain within the lower range of the observations. It can result from unresolved sources of DONI within the model such as ice-edge and under-ice phytoplankton blooms (Arrigo et al., 2012; Perrette et al., 2011), and from miss-

ing biological processes like sloppy mesozooplankton feeding and viral lysis.

In the biogeochemical model, the usable RDON, dDON, and NH_4 produced by the plankton components are taken up by bacterioplankton to build up biomass. The synthesis of cell proteins requires at least carbon and nitrogen. Bacterioplankton obtain all their carbon and some of their nitrogen from DONI (usable RDON + dDON). The simulated NH_4 uptake supplements their nitrogen requirements. The growth function is formulated using the Fasham et al. (1990) model. It assumes, in a balanced growth situation, where N and C assimilation occurs simultaneously and where bacterioplankton have fixed stoichiometry, that the ratio of NH_4 uptake to DONI uptake is constant (0.6; see Appendix A) to ensure that the biomass of the required C:N ratio is produced from DONI with a given C:N ratio. If there is not enough NH_4 available, the uptake rate of both DONI and NH_4 decreases, allowing both N and energy limitation. In Arctic waters, the inhibition of DOC uptake by bacterioplankton under inorganic nitrogen limitation was shown by Thingstad et al. (2008). However, as DONI in the model is made a proxy of DOC, the C:N ratio of the substrate is assumed constant. As a consequence, any explicit stoichiometric treatment of the simulated bacterioplankton metabolism is precluded as well as any stoichiometric coupling between DOC and inorganic nutrients (e.g. Thingstad et al., 2008). In addition, the implicit treatment of DOC in the model implies that all of the DOC required for growth is in N-containing forms. Hence it assumes that bacterioplankton cannot be N-limited in substrate. However, N-limitation of bacterioplankton production was observed in summer in surface waters of the Beaufort Sea (Ortega-Retuerta et al., 2012b). This pattern contrasts with the organic carbon limitation observed in the Yenisei and Mackenzie river plumes and the adjacent Kara and Beaufort seas (Meon and Amon, 2004; Vallières et al., 2008), hence highlighting the difficulty in drawing a general pattern on the AO scale. Nevertheless, making the C:N ratio of substrates of terrigenous and marine origin vary in a realistic way in biogeochemical models would further be required. Single explicit pools of DOC and DON represented as two different state variables, as well as a distinction between readily usable molecules (turnover within days) and more complex ones (turnover within a month) would also make the model more realistic. The parameterization of variable C:N ratios is not trivial as it requires large in situ data sets (see Letscher et al., 2015) and, in Arctic river-influenced shelf seas, a good knowledge of the characteristics of the terrigenous dissolved organic matter flowing into the coastal ocean (e.g. Mann et al., 2012). Appropriate values for the maximum uptake rates and half-saturation constants may not be easily obtained from existing data in the Arctic. As a result, the coupled model that is used in the present study is an interesting compromise relative to more complex (in terms of number of biological equations and parameters) models of

bacterioplankton growth applied to shelf waters (e.g. Auger et al., 2011; Anderson and Williams, 1998).

In the model, bacterioplankton compete with phytoplankton for the NH_4 remineralized from the usable RDON and dDON pools. This competition for a nutrient resource acts as a bottom-up control of the simulated phytoplankton and bacterioplankton production and, finally, of the BP : PP ratio. For bacterioplankton, the maximum growth rate is temperature normalized. At 10 °C, which corresponds to a sea surface temperature within the upper range of observations over the shelves in summer, it takes a value of 3 d⁻¹. Hence the NH_4 uptake efficiency (α = maximum growth rate/half-saturation constant for uptake) can reach 30 m³ mmol N⁻¹ d⁻¹, which is about 2 times higher than for small phytoplankton (14 m³ mmol N⁻¹ d⁻¹). In their study, Lignell et al. (2013) report values of α that are about 10 times higher for bacterioplankton than for small phytoplankton, hence a larger difference compared to the model. Nevertheless, the difference in α in the model is comparable to that estimated for Isefjord at the entrance of the Baltic Sea (see Lignell et al., 2013). Although the model parameterization can be improved in that respect, the model is in fair agreement with the theoretical and empirical results showing that smaller cells are more efficient in nutrient uptake than larger ones (Lignell et al., 2013). In contrast to bacterioplankton, phytoplankton uptake of inorganic nutrients is also limited by light. In the model, the diffuse attenuation of the incident light caused by the pool of coloured dissolved organic matter (0.05 m⁻¹) is set as constant in the model. This results in the light attenuation in the water column being the same in river plumes as in open and clearer waters. However, river plumes transfer to the coastal marine environment large amounts of optically active coloured dissolved organic matter of terrigenous origin that strongly attenuate the incident light propagation with depth. As the model does not account for the stronger light attenuation in river plumes, it may overestimate the simulated phytoplankton growth on NH_4 recycled from RDON by bacterioplankton and underestimate the BP in river plumes. As a consequence, the spatial and temporal evolution of the simulated BP : PP ratio can be impacted on shelves. In addition, the ability of Arctic phytoplankton to assimilate low molecular weight DON compounds (50 % of total nitrogen assimilated annually; see Simpson et al., 2013) is likely to also play an important role in the phytoplankton–bacterioplankton competition on shelves. A more accurate representation of the simulated underwater light field and uptake of nutrients in river plumes in the coupled model will certainly improve its ability to simulate the competition for nutrients between phytoplankton and bacterioplankton, and hence predict the temporal evolution of the BP : PP ratio within Arctic waters.

5 Conclusions

A pan-Arctic physical–biogeochemical model was used to quantify the contribution of usable dissolved organic nitrogen drained by the major pan-Arctic rivers to marine bacterioplankton and phytoplankton production in a scenario of melting sea ice (1998–2011). By accounting for the removal of RDON by bacterioplankton in the coupled model, the ability to predict PP and BP in river-influenced shelves is improved. The key points of the study are that

1. on average between 1998 and 2011, the removal of usable RDON by bacterioplankton is responsible for an increase of ~ 26 % in the annual BP, and an increase of ~ 8 % in the total annual PP;
2. recycled ammonium is responsible for the total PP increase; total summertime PP is increased by ~ 18 %, on average, over 1998–2011; and that
3. the processing of usable RDON by bacterioplankton promotes a higher annual BP and PP, but there is no significant temporal trend in the BP : PP ratio over 1998–2011 on the ice-free shelves; this suggests no significant evolution in the balance between autotrophy and heterotrophy in the last decade, with a constant annual flux of RDON into the coastal ocean.

The effect of the predicted warming on the Arctic watersheds is linked to a potential regional increase in RDON inputs into the AO shelf by 32–53 % before the end of the century (Frey et al., 2007). Combined with the accelerated sea-ice decline (Comiso et al., 2008) and an increase in seawater temperature on Arctic shelves (Timmermans et al., 2014), this new biogeochemical and physical setting might exacerbate the competing effect for resources between autotrophs and heterotrophs as sea ice recedes in summer. As a consequence, the metabolic state of the AO shelves could be altered. Nevertheless, to obtain robust predictions of the response of the microbial food web functioning and mass fluxes, coupled models would require improvements in parameterized land–ocean fluxes in terms of spatial and temporal variability of freshwater discharge and nutrient fluxes. In their study combining in situ data sets and modelling, Holmes et al. (2011) show that annual fluxes of RDOC in the Lena River, estimated between 1999 and 2008, can vary by about a factor of 2. Such variations accentuate the significance of considering the short-term and inter-annual variability of the continental fluxes into the coastal ocean when deriving temporal trends in plankton production and investigating potential changes in trends related to the Arctic warming. Finally, model predictions of future trajectories of PP (e.g. Vancoppenolle et al., 2013) would probably benefit from considering riverine nutrient fluxes as important drivers of PP on Arctic shelves in future decades.

However, models that are mechanistically more robust and allow for flexible stoichiometry and N-limitation of bacterial substrate uptake are probably needed for forecasting AO ecosystem responses to climate change scenarios.

Appendix A

The set of differential equations that include the mechanistic formulations cited below is given in Table 1. The biological parameters related to the mathematical equations are detailed in Table 2.

A1 Phytoplankton

The growth rate ($\mu^{\text{LP,SP}}$, d^{-1}) of large and small phytoplankton (LP and SP, respectively) depends on both light and nitrogen availability. It is computed according to Liebig's law of the minimum between the nutrient-based and light-based growth rates ($\mu_{\text{N}}^{\text{LP,SP}}$ and $\mu_{\text{light}}^{\text{LP,SP}}$, respectively):

$$\mu^{\text{LP,SP}} = \left(\mu_{\text{N}}^{\text{LP,SP}}, \mu_{\text{light}}^{\text{LP,SP}} \right). \quad (\text{A1})$$

The nutrient-based growth rate is computed as follows:

$$\mu_{\text{N}}^{\text{LP,SP}} = \mu_{\text{max}}^{\text{LP,SP}} \lim_{\text{N}}^{\text{LP,SP}}, \quad (\text{A2})$$

where $\mu_{\text{max}}^{\text{LP,SP}}$ is the maximum growth rate and $\lim_{\text{N}}^{\text{LP,SP}}$ the total nutrient limitation term (dimensionless) computed according to the substitutable model of O'Neill et al. (1989):

$$\lim_{\text{N}}^{\text{LP,SP}} = \frac{\text{NO}_3 K_{\text{NH}_4}^{\text{LP,SP}} + \text{NH}_4 K_{\text{NO}_3}^{\text{LP,SP}}}{\text{NO}_3 K_{\text{NH}_4}^{\text{LP,SP}} + \text{NH}_4 K_{\text{NO}_3}^{\text{LP,SP}} + K_{\text{NH}_4}^{\text{LP,SP}} K_{\text{NO}_3}^{\text{LP,SP}}}, \quad (\text{A3})$$

$$\lim_{\text{NO}_3}^{\text{LP,SP}} = \frac{\text{NO}_3 K_{\text{NH}_4}^{\text{LP,SP}}}{\text{NO}_3 K_{\text{NH}_4}^{\text{LP,SP}} + \text{NH}_4 K_{\text{NO}_3}^{\text{LP,SP}}}, \quad (\text{A4})$$

$$\lim_{\text{NH}_4}^{\text{LP,SP}} = \frac{\text{NH}_4 K_{\text{NO}_3}^{\text{LP,SP}}}{\text{NO}_3 K_{\text{NH}_4}^{\text{LP,SP}} + \text{NH}_4 K_{\text{NO}_3}^{\text{LP,SP}}}, \quad (\text{A5})$$

where $\lim_{\text{NO}_3}^{\text{LP,SP}}$ and $\lim_{\text{NH}_4}^{\text{LP,SP}}$ are the nitrate (NO_3) and ammonium (NH_4) uptake fractions (dimensionless), respectively. $K_{\text{NH}_4}^{\text{LP,SP}}$ and $K_{\text{NO}_3}^{\text{LP,SP}}$ are the half-saturation constants for NH_4 and NO_3 uptake, respectively. NH_4 is set to be the preferred inorganic nitrogen source (Dorch, 1990) with a higher affinity for SP (Tremblay et al., 2000). This is expressed in the model by half-saturation constants for NH_4 uptake ($K_{\text{NH}_4}^{\text{LP,SP}}$) lower than for NO_3 that, when used with the substitutable model, allow for an inhibitory effect of NH_4 on NO_3 uptake as often observed (Dorch, 1990). It implies that NO_3 uptake by LP and SP is inhibited by NH_4 at concentrations 2-fold and 10-fold lower than NO_3 concentrations, respectively. The equation used to compute the light-based growth rate is

$$\mu_{\text{light}}^{\text{LP,SP}} = \mu_{\text{max}}^{\text{LP,SP}} \lim_{\text{light}}^{\text{LP,SP}}, \quad (\text{A6})$$

where $\lim_{\text{light}}^{\text{LP,SP}}$ is the light limitation term (dimensionless) expressed as

$$\lim_{\text{light}}^{\text{LP,SP}} = 1 - e^{-\frac{E_z}{E_k^{\text{LP,SP}}}}, \quad (\text{A7})$$

where $E_k^{\text{LP,SP}}$ is the light saturation parameter ($\text{Ein m}^{-2} \text{d}^{-1}$) computed as follows:

$$E_k^{\text{LP,SP}} = \left(\frac{\text{C}}{\text{Chl}} \right)^{\text{LP,SP}} \frac{\mu_{\text{max}}^{\text{LP,SP}}}{\alpha_{\text{LP,SP}}}, \quad (\text{A8})$$

where C:Chl is the carbon to Chl ratio (g g^{-1}) and $\alpha_{\text{LP,SP}}$ is the initial slope ($\text{mg C (mg Chl)}^{-1} (\text{Ein m}^{-2} \text{d}^{-1})^{-1}$) of the photosynthesis–irradiance curve. Photoacclimation translates the adaptive response through varying C:Chl ratios in response to light and nutrient availability (e.g. Cloern et al., 1995; Geider et al., 1997; MacIntyre et al., 2002).

Varying C:Chl ratios are computed using a modified version of the empirical relationship of Cloern et al. (1995) successfully applied to Hudson Bay in the Arctic (Sibert et al., 2011). The ratios can vary up to 4- to 6-fold based on the general photoacclimation rule given by MacIntyre et al. (2002) and on Arctic nano- and pico-phytoplankton data (DuRand et al., 2002; Sherr et al., 2003) as follows:

$$\left(\frac{\text{Chl}}{\text{C}} \right)^{\text{LP}} = \left(\frac{\text{Chl}}{\text{C}} \right)_{\text{max}}^{\text{LP}} \left(1 + 4e^{-0.5 \frac{E_z}{K_E^{\text{LP}}} \lim_{\text{N}}^{\text{LP}}} \right), \quad (\text{A9})$$

$$\left(\frac{\text{Chl}}{\text{C}} \right)^{\text{SP}} = \left(\frac{\text{Chl}}{\text{C}} \right)_{\text{max}}^{\text{SP}} \left(1 + 5e^{-0.5 \frac{E_z}{K_E^{\text{SP}}} \lim_{\text{N}}^{\text{SP}}} \right), \quad (\text{A10})$$

where $K_E^{\text{LP,SP}}$ is the half-saturation parameter driving the curvature of the C:Chl versus light relationship. E_z ($\text{Ein m}^{-2} \text{d}^{-1}$) is the downwelling photosynthetically active radiation (PAR) propagating within the water column according to the Beer–Lambert law:

$$E_z = \text{PAR}_0 \int e^{-[(k_{\text{chl}} + k_w + k_{\text{nonchl}})z]} dz, \quad (\text{A11})$$

where PAR_0 is the PAR just below the sea surface. The diffuse attenuation of PAR with depth (z) is due to the simulated Chl (k_{chl}) (m^{-1} ; Morel, 1988), water molecules (k_w ; 0.04 m^{-1} ; Morel, 1988), and non-chlorophyllous matter (k_{nonchl}). k_{nonchl} is set to 0.05 m^{-1} . k_{chl} is calculated according to Morel et al. (1988) as follows:

$$k_{\text{chl}} = 0.0518 \text{Chl}^{-0.572} \text{Chl}. \quad (\text{A12})$$

$$\text{Chl} = 12 \left(\frac{106}{16} \right) \left[\text{LP} \left(\frac{\text{Chl}}{\text{C}} \right)^{\text{LP}} + \text{SP} \left(\frac{\text{Chl}}{\text{C}} \right)^{\text{SP}} \right]. \quad (\text{A13})$$

Apart from grazing, phytoplankton loss terms include senescence and sinking for LP. LP sinking rates vary in the model from 0 to 2 m d^{-1} , depending on nutrient availability (Bienfang et al., 1983):

$$\text{sed}_{\text{lp}} = \text{sed}_{\text{lp}} (1 - \lim_{\text{N}}^{\text{LP}}), \quad (\text{A14})$$

where sed_{lp} is a constant.

A2 Zooplankton

Mathematical formulations and parameters related to large zooplankton (LZ) dynamics were chosen to primarily reflect mesozooplankton. Grazing (G_{LZ} , d^{-1}) is described by an Ivlev function:

$$G_{LZ} = G_{LZ}^{\max} \left[\left(1 - e^{-\lambda(LP+SZ)} \right) \right]. \quad (A15)$$

LZ graze upon LP and protozooplankton (SZ) with a prey-specific grazing rate assumed to be proportional to the relative biomass of the prey (Campbell et al., 2009) defined for LP as follows:

$$pf_{LP} = \frac{LP}{LP + SZ}. \quad (A16)$$

Losses in LZ biomass are due to NH_4 release, fecal pellet production (non-assimilated nitrogen ingested), and mortality. Mortality encompasses senescence and predation (Eiane et al., 2002). It is described by a density-dependent quadratic function. It implicitly represents cannibalism as well as predation by macrozooplankton (Forest et al., 2012; Berline et al., 2008) and limits the occurrence of oscillations generated in such non-linear systems (Edwards and Bees, 2001). The constant of mortality is set to $0.2 \text{ (mmol N m}^{-3}\text{)}^{-1}$ to simulate realistic mortality rates (e.g. Ohman et al., 2004).

SZ grazing (G_{SZ}) upon SP and bacterioplankton (BACT) is formulated by a sigmoidal Holling-type-III function:

$$G_{SZ} = G_{SZ}^{\max} \frac{(SP + BACT)^2}{(SP + BACT)^2 + K_G^2}, \quad (A17)$$

where G_{SZ}^{\max} and K_G are the maximum grazing rate (d^{-1}) and the half-saturation constant for grazing (mmol N m^{-3}), respectively. The grazing function provides a threshold-like limit for low SP biomass that enhances the biological system stability (e.g. Steele and Henderson, 1992). In polar waters, there is evidence that protozooplankton only exert a control on small phytoplankton biomass beyond a threshold (Lancelot et al., 1997). As for LZ, SZ graze upon both SP and BACT, with a prey-specific grazing rate (d^{-1}) assumed to be proportional to the relative biomass of the prey defined for SP as follows:

$$pf_{SP} = \frac{SP}{SP + BACT}. \quad (A18)$$

We set the fraction of food ingested and converted into biomass to 30% (Straile, 1997). Lehrter et al. (1999) report that > 30% of the total nitrogen release by SZ could be in the dissolved organic form. In the model, assuming that 40% is released as detrital DON (dDON), the 60% remaining are lost as NH_4 . Other SZ loss terms are grazing by LZ and mortality. Similarly to LZ, SZ mortality is expressed by a density-dependent quadratic function to encompass grazing amongst SZ.

A3 Bacterioplankton

Bacterioplankton is explicitly simulated following the model of Fasham et al. (1990). They grow on NH_4 , dDON and usable RDON. Usable RDON is considered as 15% of total RDON (e.g. Wickland et al., 2012) and is converted into N currency (RDON) using a C : N ratio of 40 (Tank et al., 2012; Köhler et al., 2003). Dissolved organic matter (DOM) is a complex bacterial substrate representing a source of nitrogen (DON) and carbon (DOC). As nitrogen is the sole currency of the model, the simulated DONI is made a proxy of DOC for bacterioplankton uptake. This means that bacterioplankton in the model obtain all of their carbon and some of their nitrogen from the usable fraction of RDON and from detrital DON (dDON). This assumes that all of the DOC required for growth is in N-containing forms. By contrast, the simulated ammonium uptake supplements the bacterioplankton N requirements for growth. The DONI uptake rate ($U_{bactDONI}$, d^{-1}) is represented by a Michaelis–Menten model:

$$U_{bactDONI} = U_{bactmax}^{BACT} \left(\frac{DONI}{K_{DONI}^{BACT} + S + DONI} \right) Q_{10}, \quad (A19)$$

where K_{DONI}^{BACT} is the half-saturation constant for DONI uptake (mmol N m^{-3}), and S the total nitrogenous substrate (mmol N m^{-3}) defined as

$$S = (NH_4, 0.6DONI). \quad (A20)$$

According to the study by Bendtsen et al. (2002) in the Greenland Sea, a Q_{10} function was introduced using a Q_{10} factor of 3 (Kirchman et al., 2005):

$$Q_{10} = 3^{\frac{T}{10}}, \quad (A21)$$

where T is the simulated seawater temperature. A temperature normalized maximum uptake rate ($U_{bactmax}$) of $1 d^{-1}$ was used to simulate maximum growth rates in line with those measured in polar waters (e.g. Nedwell and Rutter, 1994). A growth efficiency of 20% (Ortega-Retuerta et al., 2012a; Meon and Amon, 2004) was imposed. The fractioning of the two DONI pools (i.e. usable RDON and dDON) is set as follows:

$$pf_{dDON} = \frac{dDON}{dDON + RDON}. \quad (A22)$$

Similarly, the uptake rate of NH_4 (U_{bactNH_4} , d^{-1}) is represented as follows:

$$U_{bactNH_4} = U_{bactmax}^{BACT} \left(\frac{S}{K_{NH_4}^{BACT} + S + DONI} \right) Q_{10}, \quad (A23)$$

where $K_{NH_4}^{BACT}$ is the half-saturation constant for NH_4 uptake (mmol N m^{-3}). This formulation ensures that the uptake of NH_4 will be 0.6-fold the uptake of DONI, as required by the balanced growth model (e.g. Fasham et al., 1990). BACT

cannot grow on NO_3 in the model. NO_3 uptake is an energetically expensive process, so that bacterioplankton usually accounts for more ammonium than nitrate uptake (Lipschultz, 1995). Furthermore, although a substantial nitrate uptake by bacterioplankton was reported at high latitudes, it occurred under very specific conditions such as in high-nitrate low-chlorophyll waters (Kirchman and Wheeler, 1998) or in low-chlorophyll waters dominated by cyanobacteria (Fouilland et al., 2007). Such conditions are not achieved in the model. Senescence is in the NH_4 form, and it represents 5 % of the biomass.

A4 Detritus

The pool of detrital particulate organic nitrogen (dPON) is fuelled by the production of fecal pellets by LZ, and by LZ and LP mortality. The sedimentation loss term (d^{-1}) is expressed as a quadratic function allowing for the implicit increasing aggregation of particles with increasing dPON concentration (see Guidi et al., 2008):

$$\text{sed}_{\text{pon}} = \text{sed}_{\text{dponPON}}, \quad (\text{A24})$$

where sed_{dpon} is the sedimentation constant ($\text{m d}^{-1} (\text{mmol N m}^{-3})^{-1}$). The second loss term is the dPON fragmentation into dDON (e.g. Grossart and Ploug, 2001).

The dDON pool results from dPON fragmentation, SP, SZ, and BACT mortality and SZ release. It is explicitly remineralized into NH_4 by BACT.

A5 Nutrients

NH_4 resulting from the remineralization by BACT and from the LZ and SZ release fuels regenerated primary production and BACT production. In turn, NH_4 undergoes nitrification (d^{-1}) into NO_3 as follows:

$$\text{nitrif} = \text{nitrif}_{\text{max}} \left(\frac{\text{NH}_4}{\text{NH}_4 + K_{\text{nitrif}}^{\text{N}}} \right) \left(1 - \frac{E_z}{E_z + K_{\text{nitrif}}^{\text{light}}} \right), \quad (\text{A25})$$

where $\text{nitrif}_{\text{max}}$ is the maximum nitrification rate (d^{-1}), and $K_{\text{nitrif}}^{\text{N}}$ and $K_{\text{nitrif}}^{\text{light}}$ the half-saturation constants for NH_4 (mmol N m^{-3}) and light ($\text{Ein m}^{-2} \text{d}^{-1}$) use, respectively. The latter is defined as a fraction of surface PAR (E_0) as follows:

$$K_{\text{nitrif}}^{\text{light}} = 0.005 E_0. \quad (\text{A26})$$

Mean values taken from the literature (Guerrero and Jones, 1996; Olson, 1981a, 1981b) are used to set parameters.

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References

- Adcroft, A., Hill, C., and Marshall, J.: Representation of topography by shaved cells in a height coordinate ocean model, *Mon. Weather Rev.*, 125, 2293–2315, 1997.
- Anderson, T. R and Williams, P. J. L.: Modelling the seasonal cycle of dissolved organic carbon at Station E1 in the English Channel, *Estuar. Coast. Shelf Sci.*, 46, 93–109, 1998.
- Arakawa, A. and Lamb, V.: Computational design of the basic dynamical processes of the UCLA general circulation model, *Methods Comput. Phys.*, 17, 174–267, 1977.
- Arctic Climate Impact Assessment: Arctic Climate Impact Assessment, Cambridge Press Univ., New York, 1042 pp., 2005.
- Ardyna, M., Babin, M., Gosselin, M., Devred, E., Rainville, L., and Tremblay, J.-É.: Recent Arctic Ocean sea ice loss triggers novel fall phytoplankton blooms, *Geophys. Res. Lett.*, 41, 6207–6212, doi:10.1002/2014GL061047, 2014.
- Arrigo, K. R. and van Dijken, G. L.: Secular trends in Arctic Ocean net primary production, *J. Geophys. Res.*, 116, C09011, doi:10.1029/2011JC007151, 2011.
- Arrigo, K. R., Perovich, D. K., Pickart, R. S., Brown, Z. W., van Dijken, G. L., Lowry, K. E., Mills, M. M., Palmer, M. A., Balch, W. M., Bahr, F., Bates, N. R., Benitez-Nelson, C., Bowler, B., Brownlee, E., Ehn, J. K., Frey, K. E., Garley, R., Laney, S. R., Lubelczyk, L., Mathis, J., Matsuoka, A., Mitchell, B. G., Moore, G. W., Ortega-Retuerta, E., Pal, S., Polashenski, C. M., Reynolds, R. A., Schieber, B., Sosik, H. M., Stephens, M., and Swift, J. H.: Massive phytoplankton blooms under Arctic sea ice, *Science*, 336, 1408, doi:10.1126/science.1215065, 2012.
- Auger, P. A., Diaz, F., Ulses, C., Estournel, C., Neveux, J., Joux, F., Pujo-Pay, M., and Naudin, J. J.: Functioning of the planktonic ecosystem on the Gulf of Lions shelf (NW Mediterranean) during spring and its impact on the carbon deposition: a field data and 3-D modelling combined approach, *Biogeosciences*, 8, 3231–3261, doi:10.5194/bg-8-3231-2011, 2011.
- Bélanger, S., Babin, M., and Tremblay, J.-É.: Increasing cloudiness in Arctic damps the increase in phytoplankton primary production due to sea ice receding, *Biogeosciences*, 10, 4087–4101, doi:10.5194/bg-10-4087-2013, 2013.
- Bendsten, J., Lundsgaard, C., Middelboe, M., and Archer, D.: Influence of bacterial uptake on deep-ocean dissolved organic carbon, *Global Biogeochem. Cy.*, 16, 1127, doi:10.1029/2002GB001947, 2002.
- Berline L., Spitz, Y. H., Ashjian, C. J., Campbell, R. G., Maslowski, W., and Moore, S. E.: Euphausiid transport in the Western Arctic Ocean, *Mar. Ecol.-Prog. Ser.*, 360, 163–178, 2008.
- Bienfang, P., Szyper, J., and Laws, E.: Sinking rate and pigment responses to light limitation of a marine diatom: implications to dynamics of chlorophyll maximum layers, *Oceanol. Acta*, 6, 55–62, 1983.
- Campbell, R. G., Sherr, E. B., Ashjian, C. J., Plourde, S., Sherr, B. F., Hill, V., and Stockwell, D. A.: Mesozooplankton prey preference and grazing impact in the western Arctic Ocean, *Deep-Sea Res. Pt. II*, 56, 1274–1289, doi:10.1016/j.dsr2.2008.10.027, 2009.
- Charria, G., Dadou, I., Llido, J., Dréville, M., and Garçon, V.: Importance of dissolved organic nitrogen in the north Atlantic Ocean in sustaining primary production: a 3-D modelling approach, *Biogeosciences*, 5, 1437–1455, doi:10.5194/bg-5-1437-2008, 2008.
- Cloern, J. E., Grenz, C., and Videgar-Lucas, L.: An empirical model of the phytoplankton chlorophyll:carbon ratio-the conversion factor between productivity and growth rate, *Limnol. Oceanogr.*, 40, 1313–1321, 1995.
- Codispoti, L. A., Kelly, V., Thessen, A., Matrai, P., Hill, V., Steele, M., and Light, B.: Synthesis of primary production in the Arctic Ocean: III. Nitrate and phosphate based estimates of net community production, *Prog. Oceanogr.*, 110, 126–150, doi:10.1016/j.pocean.2012.11.006, 2013.
- Condran, A., Winsor, P., Hill, C. N., and Menemenlis, D.: Response of Arctic freshwater budget to extreme NAO forcing, *J. Clim.*, 22, 2422–2437, 2009.
- Comiso, J. C., Parkinson, C. L., Gersten, R., and Stock, L.: Accelerated decline in the Arctic sea ice cover, *Geophys. Res. Lett.*, 35, L01703, doi:10.1029/2007GL031972, 2008.
- Coupel, P., Jin, H. Y., Joo, M., Horner, R., Bouvet, H. A., Sicre, M.-A., Gascard, J.-C., Chen, J. F., Garçon, V., and Ruiz-Pino, D.: Phytoplankton distribution in unusually low sea ice cover over the Pacific Arctic, *Biogeosciences*, 9, 4835–4850, doi:10.5194/bg-9-4835-2012, 2012.
- Dorch, Q.: The interaction between ammonium and nitrate uptake in phytoplankton, *Mar. Ecol.-Prog. Ser.*, 61, 183–201, 1990.
- Ducklow, H. W.: The bacterial component of the oceanic euphotic zone, *FEMS Microbiol. Ecol.*, 30, 1–10, 1999.
- DuRand, M. D., Green, R. E., Sosik, H. M., and Olson, R. J.: Diel variations in Optical properties of *Micromonas pusilla* (Prasinophyceae), *J. Phycol.*, 38, 1132–1142, 2002.
- Edwards, A. M. and Bees, M. A.: Generic dynamics of a simple plankton population model with a non-integer exponent of closure, *Chaos Solitons Fractals*, 12, 289–300, 2001.
- Eiane, K., Aksnes, D. L., Ohman, M. D., Wood, S., and Martnussen, M. B.: Stage-specific mortality of *Calanus* spp. under different predation regimes, *Limnol. Oceanogr.*, 47, 636–645, 2002.
- Fasham, M. J. R., Ducklow, H. W., and McKelvie, S. M.: A nitrogen-based model of plankton dynamics in the oceanic mixed layer, *J. Mar. Res.*, 48, 591–639, 1990.
- Forest, A., Stemmann, L., Picheral, M., Burdorf, L., Robert, D., Fortier, L., and Babin, M.: Size distribution of particles and zooplankton across the shelf-basin system in southeast Beaufort Sea: combined results from an Underwater Vision Profiler and vertical net tows, *Biogeosciences*, 9, 1301–1320, doi:10.5194/bg-9-1301-2012, 2012.

- Fouilland, E., Gosselin, M., Rivkin, R. B., Vasseur, C., and Mostajir, B.: Nitrogen uptake by heterotrophic bacteria and phytoplankton in Arctic surface waters, *J. Plankton Res.*, 29, 369–376, 2007.
- Frey, K. E., McClelland, J. W., Holmes, R. M., and Smith, L. C.: Impacts of climate warming and permafrost thaw on the riverine transport of nitrogen and phosphorus to the Kara Sea, *J. Geophys. Res.*, 112, G04S58, doi:10.1029/2006JG000369, 2007.
- Garneau, M. E., Roy, S., Lovejoy, C., Gratton, Y., and Vincent, W. F.: Seasonal dynamics of bacterial biomass and production in a coastal arctic ecosystem: Franklin Bay, western Canadian Arctic, *J. Geophys. Res.*, 113, C07S91, doi:10.1029/2007JC004281, 2008.
- Geider, R. J., MacIntyre, H. L., and Kana, T. M.: Dynamic model of phytoplankton growth and acclimation: responses of the balanced growth rate and the chlorophyll a: carbon ratio to light, nutrient-limitation and temperature, *Mar. Ecol.-Progr. Ser.*, 148, 187–200, 1997.
- Guerrero, M. A. and Jones, R. D.: Photoinhibition of marine nitrifying bacteria, II. Dark recovery after monochromatic or polychromatic irradiation, *Mar. Ecol.-Prog. Ser.*, 141, 193–198, 1996.
- Grossart, H.-P. and Ploug, H.: Microbial degradation of organic carbon and nitrogen on diatom aggregates, *Limnol. Oceanogr.*, 46, 267–277, 2001.
- Guidi, L., Jackson, G. A., Stemmann, L., Miquel, J. C., Picheral, M., and Gorsky, G.: Relationship between particle size distribution and flux in the mesopelagic zone, *Deep-Sea Res. Pt. I*, 55, 1364–1374, 2008.
- Hibler III, W. D.: A dynamic thermodynamic sea ice model, *J. Phys. Oceanogr.*, 9, 815–846, 1979.
- Hibler III, W. D. and Bryan, K.: A diagnostic ice-ocean model, *J. Phys. Oceanogr.*, 17, 987–1015, 1987.
- Hill, V. J., Matrai, P., Olson, E., Suttle, S., Steele, M., Codispoti, L., and Zimmerman, R.: Synthesis of integrated primary production in the Arctic Ocean: II. In situ and remotely sensed estimates, *Progr. Oceanogr.*, 110, 107–125, doi:10.1016/j.pocean.2012.11.005, 2013.
- Holmes, R. M., McClelland, J. W., Peterson, B. J., Tank, S. E., Buliygina, E., Eglinton, T. I., Gordeev, V. V., Gurtovaya, T. Y., Raymond, P. A., Repeta, D. J., Staples, R., Stiegl, R. G., Zhulidov, A. V., and Zimov, S. A.: Seasonal and annual fluxes of nutrients and organic matter from large rivers to the Arctic Ocean and surrounding seas, *Estuar. Coast*, 35, 369–382, doi:10.1007/s12237-011-9386-6, 2011.
- Jakobsson, M., Macnab, R., Mayer, L., Anderson, R., Edwards, M., Hatzky, J., Schenke, H. W., and Johnson, P.: An improved bathymetric portrayal of the Arctic Ocean: Implications for ocean modeling and geological, geophysical and oceanographic analyses, *Geophys. Res. Lett.*, 35, L07602, doi:10.1029/2008GL033520, 2008.
- Kahru, M., Brotas, V., Manzano-Sarabia, M., and Mitchell, B. G.: Are phytoplankton blooms occurring earlier in the Arctic?, *Glob. Change Biol.*, 17, 1733–1739, doi:10.1111/j.1365-2486.2010.02312.x, 2011.
- Köhler, H., Meon, B., Gordeev, V. V., Spitzky, A. and Amon, R. M. W.: Dissolved organic matter (DOM) in the estuaries of Ob and Yenisei and the adjacent Kara Sea, Russia, in: *Siberian River Run-off in the Kara Sea: Characterisation, Quantification, Variability, and Environmental Significance*, edited by: Stein, R., Fahl, K., Fütterer, D. K., Galimov, E. M., and Stepanets, O. V., *Proceedings in Marine Sciences*, 6, Elsevier, Amsterdam, 281–308, 2003.
- Kirchman, D. L. and Wheeler, P. A.: Uptake of ammonium and nitrate by heterotrophic bacteria and phytoplankton in the sub-Arctic Pacific, *Deep-Sea Res. Pt. I*, 45, 347–365, 1998.
- Kirchman, D. L., Malmstrom, R. R., and Cottrell, M. T.: Control of bacterial growth by temperature and organic matter in the Western Arctic, *Deep-Sea Res. Pt. II*, 52, 3386–3395, doi:10.1016/j.dsr2.2005.09.005, 2005.
- Kirchman, D. L., Hill, V., Cottrell, M. T., Gradinger, R., Malmstrom, R. R., and Parker, A.: Standing stocks, production, and respiration of phytoplankton and heterotrophic bacteria in the western Arctic Ocean, *Deep-Sea Res. Pt. II*, 56, 1237–1248, doi:10.1016/j.dsr2.2008.10.018, 2009.
- Kristiansen, S., Fabrot, T., and Wheeler, P. A.: Nitrogen cycling in the Barents Sea, Seasonal dynamics of new and regenerated production in the Marginal Ice Zone, *Limnol. Oceanogr.*, 39, 1630–1642, 1994.
- Lammers, R. B., Shiklomanov, A. I., Vörösmarty, C. J., Fekete, B. M., and Peterson, B. J.: Assessment of contemporary Arctic river runoff based on observational discharge records, *J. Geophys. Res.*, 106, 3321–3334, 2001.
- Lancelot, C., Becquevort, S., Menon, P., Mathot, S., and Dandois, J.-M.: Ecological modelling of the planktonic microbial food-web, in: *Belgian Research Program on the Antarctic, Scientific Results of Phase III (1992–1996): Marine Biogeochemistry and Ecodynamics*, edited by: Caschetto, S., 1, 1–78, Fed. Off. for Sci., Tech. and Cult. Affairs, Brussels, 1997.
- Le Fouest, V., Zakardjian, B., Saucier, F. J., and Starr, M.: Seasonal versus synoptic variability in planktonic production in a high-latitude marginal sea: the Gulf of St. Lawrence (Canada), *J. Geophys. Res.*, 110, C099012, doi:10.1029/2004JC002423, 2005.
- Le Fouest, V., Zakardjian, B., Saucier, F. J., and Cizmeli, S. A.: Application of SeaWIFS- and AVHRR-derived data for mesoscale and regional validation of a 3-D high-resolution physical-biological model of the Gulf of St. Lawrence (Canada), *J. Mar. Syst.*, 60, 30–50, 2006.
- Le Fouest, V., Postlethwaite, C., Morales Maqueda, M. A., Bélanger, S., and Babin, M.: On the role of tides and strong wind events in promoting summer primary production in the Barents Sea, *Cont. Shelf Sci.*, 31, 1869–1879, doi:10.1016/j.csr.2011.08.013, 2011.
- Le Fouest, V., Babin, M., and Tremblay, J.-É.: The fate of riverine nutrients on Arctic shelves, *Biogeosciences*, 10, 3661–3677, doi:10.5194/bg-10-3661-2013, 2013a.
- Le Fouest, V., Zakardjian, B., Xie, H., Raimbault, P., Joux, F., and Babin, M.: Modeling plankton ecosystem functioning and nitrogen fluxes in the oligotrophic waters of the Beaufort Sea, Arctic Ocean: a focus on light-driven processes, *Biogeosciences*, 10, 4785–4800, doi:10.5194/bg-10-4785-2013, 2013b.
- Lehrter, J. C., Pennock, J. R., and McManus, G. B.: Microzooplankton grazing and nitrogen excretion across a surface estuarine-coastal interface, *Estuaries*, 22, 113–125, 1999.
- Letscher, R. T., Moore, J. K., Teng, Y.-C., and Primeau, F.: Variable C : N : P stoichiometry of dissolved organic matter cycling in the Community Earth System Model, *Biogeosciences*, 12, 209–221, doi:10.5194/bg-12-209-2015, 2015.

- Li, W. K. W., McLaughlin, F. A., Lovejoy, C., and Carmack, E. C.: Smallest algae thrive as the Arctic Ocean freshens, *Science*, 326, 539, doi:10.1126/science.1179798, 2009.
- Lignell, R., Haario, H., Laine, M., and Thingstad, T. F.: Getting the “right” parameter values for models of the pelagic microbial food web, *Limnol. Oceanogr.*, 58, 301–313, 2013.
- Lipschultz, F.: Nitrogen-specific uptake rates of marine phytoplankton isolated from natural populations of particles by flow cytometry, *Mar. Ecol.-Prog. Ser.*, 123, 245–258, 1995.
- Manizza, M., Follows, M. J., Dutkiewicz, S., McClelland, J. W., Menemenlis, D., Hill, C. N., Townsend-Small, A., and Peterson, B. J.: Modeling transport and fate of riverine dissolved organic carbon in the Arctic Ocean, *Global Biogeochem. Cy.*, 23, GB4006, doi:10.1029/2008GB003396, 2009.
- Manizza, M., Follows, M. J., Dutkiewicz, S., Menemenlis, D., Hill, C. N., and Key, R. M.: Changes in the Arctic Ocean CO₂ sink (1996–2007): a regional model analysis, *Global Biogeochem. Cy.*, 27, 1108–1118, doi:10.1002/2012GB004491, 2013.
- Mann, P. J., Davydova, A., Zimov, N., Spencer, R. G. M., Davydov, S., Bulygina, E., Zimov, S., and Holmes, R. M.: DOM composition and lability during the Arctic spring freshet on the River Kolyma, Northeast Siberia, *J. Geophys. Res.*, 117, G01028, doi:10.1029/2011JG001798, 2012.
- Marshall, J., Hill, C., Perelman, L., and Adcroft, A.: Hydrostatic, quasi-hydrostatic and nonhydrostatic ocean modeling, *J. Geophys. Res.*, 102, 5733–5752, 1997.
- McClelland, J. W., Holmes, R. M., Peterson, B. J., Amon, R., Brabets, T., Cooper, L., Gibson, J., Gordeev, V. V., Guay, C., Milburn, D., Staples, R., Raymond, P. A., Shiklomanov, I., Striegl, R., Zhulidov, A., Gurtovaya, T., and Zimov, S.: Development of a pan-Arctic database for river chemistry, *Eos Trans. AGU*, 89, 217–218, doi:10.1029/2008EO240001, 2008.
- MacIntyre, H. L., Kana, T. M., Anning, T., and Geider, R. J.: Photoacclimation of photosynthesis irradiance response curves and photosynthetic pigments in microalgae and cyanobacteria, *J. Phycology*, 38, 17–38, 2002.
- Menemenlis, D., Hill, C., Adcroft, A., Campin, J.-M., Cheng, B., Ciotti, B., Fukumori, I., Heimbach, P., Henze, C., Kohl, A., Lee, T., Stammer, D., Taft, J., and Zhang, J.: NASA supercomputer improves prospects for ocean climate research, *Eos Trans. AGU*, 86, 89–96, 2005.
- Meon, B., and Amon, R. M. W.: Heterotrophic bacterial activity and fluxes of dissolved free amino acids and glucose in the Arctic rivers Ob, Yenisei and the adjacent Kara Sea, *Aquat. Microb. Ecol.*, 37, 121–135, 2004.
- Morel, A.: Optical modeling of the upper ocean in relation to its biogenous matter content (case I waters), *J. Geophys. Res.*, 93, 10749–10768, 1988.
- National Oceanographic Data Centre World Ocean Atlas 2005: Documentation accompanying WOA05 DVD (ASCII text and portable document format), Prepared by the Ocean Climate Laboratory, National Oceanographic Data Center, Silver Springs, MD 20910, 12 pp., 2006.
- Nedwell, D. B. and Rutter, M.: Influence of temperature on growth rate and competition between two psychrotolerant antarctic bacteria: low temperature diminishes affinity for substrate uptake, *Appl. Environ. Microbiol.*, 60, 1984–1992, 1994.
- Ohman, M. D., Eiane, K., Durbin, E. G., Runge, J. A., and Hirche, H.-J.: A comparative study of *Calanus finmarchicus* mortality patterns at five localities in the North Atlantic, *ICES J. Mar. Sci.*, 61, 687–697, 2004.
- Olson, R. J.: Differential photoinhibition of marine nitrifying bacteria: a possible mechanism for the formation of the primary nitrite maximum, *J. Mar. Res.*, 39, 227–238, 1981a.
- Olson, R. J.: 15N tracers studies of the primary nitrite maximum, *J. Mar. Res.*, 39, 203–226, 1981b.
- O’Neill, R. V., DeAngelis, D. L., Pastor, J. J., Jackson, B. J., and Post, W. M.: Multiple nutrient limitations in ecological models, *Ecol. Model.*, 46, 147–163, 1989.
- Onogi, K., Tsutsui, J., Koide, H., Sakamoto, M., Kobayashi, S., Hatushika, H., Matsumoto, T., Yamazaki, N., Kamahori, H., Takahashi, K., Kadokura, S., Wada, K., Kato, K., Oyama, R., Ose, T., Mannoji, N., and Taira, R.: The JRA?25 reanalysis, *J. Meteorol. Soc. Jpn.*, 85, 369–432, 2007.
- Opsahl, S., Benner, R., and Amon, R. M. W.: Major flux of terrigenous dissolved organic matter through the Arctic Ocean, *Limnol. Oceanogr.*, 44, 2017–2023, 1999.
- Ortega-Retuerta, E., Jeffrey, W. H., Babin, M., Bélanger, S., Benner, R., Marie, D., Matsuoka, A., Raimbault, P., and Joux, F.: Carbon fluxes in the Canadian Arctic: patterns and drivers of bacterial abundance, production and respiration on the Beaufort Sea margin, *Biogeosciences*, 9, 3679–3692, doi:10.5194/bg-9-3679-2012, 2012a.
- Ortega-Retuerta, E., Jeffrey, W. H., Ghiglione, J. F., and Joux, F.: Evidence of heterotrophic prokaryotic activity limitation by nitrogen in the Western Arctic Ocean during summer, *Polar Biol.*, 35, 785–794, 2012b.
- Perrette, M., Yool, A., Quartly, G. D., and Popova, E. E.: Near-ubiquity of ice-edge blooms in the Arctic, *Biogeosciences*, 8, 515–524, doi:10.5194/bg-8-515-2011, 2011.
- Piontek, J., Sperling, M., Nothig, E.-M., and Engel, A.: Regulation of bacterioplankton activity in Fram Strait (Arctic Ocean) during early summer: the role of organic matter supply and temperature, *J. Mar. Syst.*, 132, 83–94, 2014.
- Rachold, V., Eiken, H., Gordeev, V. V., Grigoriev, M. N., Hubberten, H.-W., Lisitzin, A. P., Shevchenko, V. P., and Schirmeister, L.: Modern terrigenous organic carbon input to the Arctic Ocean, in: *The Organic Carbon Cycle in the Arctic Ocean*, edited by: Stein, R. S. and Macdonald, R. W., 33–55, Springer, New York, 2004.
- Raymond, P. A., McClelland, J. W., Holmes, R. M., Zhulidov, A. V., Mull, K., Peterson, B. J., Striegl, R. G., Aiken, G. R., and Gurtovaya, T. Y.: Flux and age of dissolved organic carbon exported to the Arctic Ocean: A carbon isotopic study of the five largest arctic rivers, *Global Biogeochem. Cy.*, 21, GB4011, doi:10.1029/2007GB002934, 2007.
- Redfield, A. C., Ketchum, B. H., and Richards, F. A.: The influence of organisms on the composition of sea water, in *The Sea: Ideas and Observations on Progress in the Study of the Seas*, edited by: Hill, M. N., 26–27, Wiley-Intersci., Hoboken, NY, 1973.
- Rich, J., Gosselin, M., Sherr, E., Sherr, B., and Kirchman, D. L.: High bacterial production, uptake and concentrations of dissolved organic matter in the Central Arctic Ocean, *Deep-Sea Res. Pt. II*, 44, 1645–1663, doi:10.1016/S0967-0645(97)00058-1, 1997.
- Sakshaug, E.: Primary and Secondary Production in the Arctic Seas, in: *The Organic Carbon Cycle in the Arctic Ocean*, edited by: Stein, R. and Macdonald, R. W., 57–81, doi:10.1007/978-3-642-18912-8_3, Springer Berlin Heidelberg, 2004.

- Sahzin, A. F., Romanova, N. D., and Mosharov, S. A.: Bacterial and primary production in the pelagic zone of the Kara Sea, *Oceanology*, 50, 759–765, 2010.
- Serreze, M., Barret, A. P., Slater, A. G., Woodgate, R. A., Aagard, K., Lammers, R. B., Steele, M., Mortitz, R., Meredith, M., and Lee, C. M.: The large-scale fresh water cycle of the Arctic, *J. Geophys. Res.*, 111, C11010, doi:10.1029/2005JC003424, 2006.
- Sherr, E. B., Sherr, B. F., Wheeler, P. A., and Thompson, K.: Temporal and spatial variation in stocks of autotrophic and heterotrophic microbes in the upper water column of the central Arctic Ocean, *Deep-Sea Res. Pt. I*, 50, 557–571, doi:10.1016/S0967-0637(03)00031-1, 2003.
- Shiklomanov, I., Shiklomanov, A., Lammers, R., Peterson, B., and Vorosmarty, C.: The dynamics of river water inflow to the Arctic Ocean, in *The Freshwater Budget of the Arctic Ocean*, edited by: E. Lewis, 281–296, Kluwer Acad., Dordrecht, Netherlands, 2000.
- Shiklomanov, A. and Lammers, R. B.: River discharge (in Arctic, Report Card 2011), available at http://www.arctic.noaa.gov/report11/river_discharge.html (last access: 27 October 2014), 2011.
- Sibert, V., Zakardjian, B., Gosselin, M., Starr, M., Senneville, S., and LeClainche, Y.: 3D bio-physical model of the sympagic and planktonic productions in the Hudson Bay System. *J. Mar. Syst.*, 88, 401–422, doi:10.1016/j.jmarsys.2011.03.014, 2011.
- Simpson, K. G., Tremblay, J.-É., Brugel, S., and Price, N. M.: Nutrient dynamics in the western Canadian Arctic. II. Estimates of new and regenerated production over the Mackenzie Shelf and Cape Bathurst Polynya, *Mar. Ecol.-Prog. Ser.*, 484, 47–62, doi:10.3354/meps10298, 2013.
- Steele, J. H. and Henderson, E. W.: The role of predation in plankton models, *J. Plankton Res.*, 14, 157–172, 1992.
- Steele, M., Morley, R., and Ermold, W.: PHC: A global ocean hydrography with a high quality Arctic Ocean, *J. Clim.*, 14, 2079–2087, doi:10.1175/1520-0442(2001)014<2079:PAGOHW>2.0.CO;2, 2001.
- Steele, M., Ermold, W., and Zhang, J.: Arctic Ocean surface warming trends over the past 100 years, *Geophys. Res. Letters*, 35, L02614, doi:10.1029/2007GL031651, 2008.
- Steward, G. F., Smith, D. C., and Azam, F.: Abundance and production of bacteria and viruses in the Bering and Chukchi Seas, *Mar. Ecol.-Prog. Ser.*, 131, 287–300, 1996.
- Straile, D.: Gross growth efficiencies of protozoan and metazoan zooplankton and their dependence on food concentration, predator-prey weight ratio, and taxonomic group, *Limnol. Oceanogr.*, 42, 1375–1385, 1997.
- Sturluson M., Nielsen, T. G., and Wassmann, P.: Bacteria abundance, biomass and production during spring blooms in the northern Barents Sea, *Deep-Sea Res. Pt. II*, 55, 2186–2198, 2008.
- Taniguchi, A.: Differences in structure of lower trophic levels of pelagic ecosystems in the eastern and western subarctic Pacific, *Prog. Oceanogr.*, 43, 289–315, 1999.
- Tank, S. E., Manizza, M., Holmes, R. M., McClelland, J. W., and Peterson, B. J.: The processing and impact of dissolved riverine nitrogen in the Arctic Ocean, *Estuar. Coasts*, 35, 401–415, doi:10.1007/s12237-011-9417-3, 2012.
- Thingstad, F. T., Bellerby, R. G. J., Bratbak, G., Børshheim, K. Y., Egge, J. K., Heldal, M., Larsen, A., Neill, C., Nejstgaard, J., Norland, S., Sandaa, R.-A., Skjoldal, E., Tanaka, T., Thyrhaug, R., and Töpper, B.: Counterintuitive carbon-to-nutrient coupling in an Arctic pelagic ecosystem, *Nature*, 455, 387–390, 2008.
- Timmermans, M.-L., Ashik, I., Cao, Y., Frolov, I., Ha, H. K., Ingvaldsen, R., Kikuchi, T., Kim, T. W., Krishfield, R., Loeng, H., Nishino, S., Pickart, R., Polyakov, I., Rabe, B., Semiletov, I., Schauer, U., Schlosser, P., Shakhova, N., Smethie, W. M., Sokolov, V., Steele, M., Su, J., Toole, J., Williams, W., Woodgate, R., Zhao, J., Zhong, W., and Zimmermann, S.: Arctic Ocean Sea Surface Temperature (in Arctic Report Card), available at: http://www.arctic.noaa.gov/reportcard/ocean_temperature_salinity.html (last access: 27 October, 2014), 2014.
- Tremblay, J.-É., Legendre, L., Klein, B. and Therriault, J.-C.: Size differential uptake of nitrogen and carbon in a marginal sea (Gulf of St. Lawrence, Canada): Significance of diel periodicity and urea uptake, *Deep Sea Res. II*, 47, 489–518, 2000.
- Vancoppenolle, M., Bopp, L., Madec, G., Dunne, J., Ilyina, T., Halloran, P. R., and Steiner, N.: Future Arctic Ocean primary productivity from CMIP5 simulations: Uncertain outcome, but consistent mechanisms, *Global Biogeochem. Cy.*, 27, 605–619, doi:10.1002/gbc.20055, 2013.
- Vallièeres, C., Retamal, L., Osburn, C., and Vincent, W. F.: Bacterial production and microbial food web structure in a large Arctic river and the coastal Arctic Ocean, *J. Mar. Syst.*, 74, 756–773, doi:10.1016/j.jmarsys.2007.12.002, 2008.
- Wickland, K. P., Aiken, G. R., Butler, K., Dornblaser, M. M., Spencer, R. G. M., and Striegl, R. G.: Biodegradability of dissolved organic carbon in the Yukon River and its tributaries: Seasonality and importance of inorganic nitrogen, *Global Biogeochem. Cy.*, 26, GB0E03, doi:10.1029/2012GB004342, 2012.
- Xie, H., Bélanger, S., Song, G., Benner, R., Taalba, A., Blais, M., Tremblay, J.-É., and Babin, M.: Photoproduction of ammonium in the southeastern Beaufort Sea and its biogeochemical implications, *Biogeosciences*, 9, 3047–3061, doi:10.5194/bg-9-3047-2012, 2012.
- Yang, D., Shic, X., and Marsh, P.: Variability and extreme of Mackenzie River daily discharge during 1973–2011, *Quatern. Internat.*, doi:10.1016/j.quaint.2014.09.023, in press, 2015.
- Zhang, J. and Hibler III, W. D.: On an efficient numerical method for modeling sea ice dynamics, *J. Geophys. Res.*, 102, 8691–8702, 1997.
- Zhang, J. and Rothrock, D.: Modeling global sea ice with a thickness and enthalpy distribution model in generalized curvilinear coordinates, *Mon. Weather Rev.*, 131, 845–861, doi:10.1175/1520-0493(2003)131<0845:MGSIIWA>2.0.CO;2, 2003.